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Document provides a summary of important aquatic toxicological data pertaining to mercury. Criterion Maximum and Criterion Continuous Concentrations (CMC and CCC) are calculated. Any necessary adjustments to the criterion are discussed. An extensive bibliography is provided.

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AMBIENT AQUATIC LIFE WATER QUALITY CRITERIA FOR MERCURY

U.S. ENVIRONMENTAL PROTECTION AGENCY
OFFICE OF RESEARCH AND DEVELOPMENT
ENVIRONMENTAL RESEARCH LABORATORIES
DULUTH, MINNESOTA
NARRAGANSETT, RHODE ISLAND

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FOREWORD

Section 304(a)(1) of the Clean Water Act of 1977 (P.L. 95-217) requires the Administrator of the Environmental Protection Agency to publish criteria for water quality accurately reflecting the latest scientific knowledge on the kind and extent of all identifiable effects on health and welfare which may be expected from the presence of pollutants in any body of water, including ground water. This document is a revision of proposed criteria based upon a consideration of comments received from other Federal agencies, State agencies, special interest groups, and individual scientists. The criteria contained in this document replace any previously published EPA aquatic life criteria.

The term "water quality criteria" is used in two sections of the Clean Water Act, section 304(a)(1) and section 303(c)(2). The term has a different program impact in each section. In section 304, the term represents a non-regulatory, scientific assessment of ecological effects. The criteria presented in this publication are such scientific assessments. Such water quality criteria associated with specific stream uses when adopted as State water quality standards under section 303 become enforceable maximum acceptable levels of a pollutant in ambient waters. The water quality criteria adopted in the State water quality standards could have the same numerical limits as the criteria developed under section 304. However, in many situations States may want to adjust water quality criteria developed under section 304 to reflect local environmental conditions and human exposure patterns before incorporation into water quality standards. It is not until their adoption as part of the State water quality standards that the criteria become regulatory.

Guidelines to assist the States in the modification of criteria presented in this document, in the development of water quality standards, and in other water-related programs of this Agency, have been developed by EPA.

Edwin L. Johnson
Director
Office of Water Regulations and Standards

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J. Howard McCormick
(freshwater author)
Environmental Research Laboratory
Duluth, Minnesota

John H. Gentile
(saltwater author)
Environmental Research Laboratory
Narragansett, Rhode Island

Charles E. Stephan
(document coordinator)
Environmental Research Laboratory
Duluth, Minnesota

David J. Hansen
(saltwater coordinator)
Environmental Research Laboratory
Narragansett, Rhode Island

Judy L. Crane
University of Wisconsin-Superior
Superior, Wisconsin

Clerical Support: Terry L. Highland

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Introduction*

Mercury has long been recognized as one of the most toxic of the heavy metals, but only recently was it identified as a serious pollutant in the aquatic environment (National Research Council, 1978; National Research Council Canada, 1979; Nriagu, 1979). Elemental mercury is a heavy liquid at room temperature and was considered relatively inert, because it was assumed that it would quickly settle to the bottom of a body of water and remain there in an innocuous state. However, elemental mercury is oxidized to mercury(II) under natural conditions (Wood, 1974). Furthermore, mercury(II), whether discharged directly or produced from elemental mercury, can be methylated by both aerobic and anaerobic bacteria (Akagi, 1979; Beijer and Jernelov, 1979; Callahan, et al. 1979; Jernelov, 1971, 1972; Jernelov, et al. 1975; National Research Council, 1978; Summers and Silver, 1978; Thayer and Brinckman, 1982; Wright and Hamilton, 1982). Mercury(II) can also be methylated in the slime coat, liver, and intestines of fish (Jernelov, 1968; Matsumura, et al. 1975; Rudd, et al. 1980b), but methylation apparently does not occur in other tissues (Huckabee, et al. 1978; Matida, et al. 1971; Pennacchioni, et al. 1976) or in plants (Czuba and Mortimer, 1980). (The term "methylmercury" is used herein to refer only to monomethylmercury, and not to dimethylmercury or any other monoorganomercury salt or diorganomercury compound. Inorganic mercury(II) will be referred to as "mercury(II)".)

The importance of methylation may be reduced by demethylation (Bisogni, 1979; Ramamoorthy, et al. 1982). Demethylation might provide a feedback

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*An understanding of the "Guidelines for Deriving Numerical National Water Quality Criteria for the Protection of Aquatic Organisms and Their Uses" (Stephan, et al., 1985), hereafter referred to as the Guidelines, is necessary in order to understand the following text, tables, and calculations.

mechanism that controls the concentration of methylmercury in sediment and in water. Jernelov, et al. (1975) cited a case in which low levels of methylmercury in fish from a highly contaminated area coincided with strong methylmercury degrading activity in the sediment. Demethylation also occurs in fish (Burrows and Krenkel, 1973; de Freitas, et al. 1981; Gage, 1964; Olson, et al. 1978), probably as part of the depuration mechanism.

Numerous factors such as alkalinity, ascorbic acid, chloride, dissolved oxygen, hardness, organic complexing agents, pH, sediment, and temperature probably affect the acute and chronic toxicity and bioaccumulation of the various forms of mercury (Amend, et al. 1969; Baker, et al. 1983; Feich, et al. 1972; Hahne and Kroontje, 1973; Jernelov, 1980; Ramamoorthy and Blumhagen, 1984; Ribeyre and Boudou, 1982; Rogers and Beamish, 1981, 1983; Rudd, et al. 1980a; Rudd and Turner, 1983a,b; Sharma, et al. 1982; Stokes, et al. 1983; Tsai, et al. 1975; Wren and MacCrimmon, 1983; Wright and Hamilton, 1982).

A variety of studies have been conducted on the effect of selenium on the acute toxicity of mercury (e.g., Birge, et al. 1981; Bowers, et al. 1980; De Filippis, 1979; Heisinger, 1981; Heisinger, et al. 1979; Klaverkamp, et al., 1983a; Lawrence and Holoka, 1983; Sharma and Davis, 1980c) and on the accumulation of mercury from food and water (e.g., Beijer and Jernelov, 1978; Chang, et al. 1983; Heisinger, et al. 1979; Klaverkamp, et al. 1983b; Rudd, et al. 1980a; Sharma and Davis, 1980c; Speyer, 1980; Turner and Swick, 1983). Available data do not, however, show that quantitative relationships are consistent enough for a variety of aquatic species to enable relating water quality criteria to any of these variables.

Because of the variety of forms of inorganic and organic mercury and lack of definitive information about their relative toxicities, no available

analytical measurement is known to be ideal for expressing aquatic life criteria for mercury. Previous aquatic life criteria for mercury (U.S. EPA, 1980) were specified in terms of total recoverable mercury, which would probably be measured as total mercury (U.S. EPA, 1983a), but both of these measurements are probably too rigorous in some situations. Acid-soluble mercury (operationally defined as the mercury that passes through a 0.45 μm membrane filter after the sample is acidified to pH = 1.5 to 2.0 with nitric acid) is probably the best measurement at the present for the following reasons:

1. This measurement is compatible with all available data concerning toxicity of mercury to, and bioaccumulation of mercury by, aquatic organisms. No test results were rejected just because it was likely that they would have been substantially different if they had been reported in terms of acid-soluble mercury. For example, results reported in terms of dissolved mercury would not have been used if the concentration of precipitated mercury was substantial.
2. On samples of ambient water, measurement of acid-soluble mercury should measure all forms of mercury that are toxic to aquatic life or can be readily converted to toxic forms under natural conditions. In addition, this measurement should not measure several forms, such as mercury that is occluded in minerals, clays, and sand or is strongly sorbed to particulate matter, that are not toxic and are not likely to become toxic under natural conditions. Although this measurement (and many others) will measure soluble, complexed forms of mercury, such as the EDTA complex of mercury(II), that probably have low toxicities to aquatic life, concentrations of these forms probably are negligible in most ambient water.

3. Although water quality criteria apply to ambient water, the measurement used to express criteria is likely to be used to measure mercury in aqueous effluents. Measurement of acid-soluble mercury should be applicable to effluents because it will measure precipitates, such as carbonate and hydroxide precipitates of mercury(II), that might exist in an effluent and dissolve when the effluent is diluted with receiving water. If desired, dilution of effluent with receiving water before measurement of acid-soluble mercury might be used to determine whether the receiving water can decrease the concentration of acid-soluble mercury because of sorption.
4. The acid-soluble measurement should be useful for most metals, thus minimizing the number of samples and procedures that are necessary.
5. The acid-soluble measurement does not require filtration at the time of collection, as does the dissolved measurement.
6. The only treatment required at the time of collection is preservation by acidification to pH = 1.5 to 2.0, similar to that required for the total measurement.
7. Durations of 10 minutes to 24 hours between acidification and filtration probably will not affect the result substantially.
8. The carbonate system has a much higher buffer capacity from pH = 1.5 to 2.0 than it does from pH = 4 to 9 (Weber and Stumm, 1963).
9. Differences in pH within the range of 1.5 to 2.0 probably will not affect the result substantially.
10. After acidification and filtration of the sample to isolate the acid-soluble mercury, the analysis for total acid-soluble mercury can be performed using permanganate and persulfate oxidation and cold vapor atomic absorption (U.S. EPA, 1983a), as with the total measurement.

Acid-soluble inorganic mercury can be measured by not breaking down the organomercury compounds before using cold vapor atomic absorption. Methylmercury has been measured using gas chromatography (Cappon, 1984; Hildebrand, et al. 1980; Paasivirta, et al. 1981), thin layer chromatography (Kudo, et al. 1982), and liquid chromatography (Gast and Kraak, 1979; MacCrehan and Durst, 1978).

Thus, expressing aquatic life criteria for mercury in terms of the acid-soluble measurement has both toxicological and practical advantages. On the other hand, because no measurement is known to be ideal for expressing aquatic life criteria for mercury or for measuring mercury in ambient water or aqueous effluents, measurement of both total acid-soluble mercury and total mercury in ambient water or effluent or both might be useful. For example, there might be cause for concern if total mercury is much above an applicable limit, even though total acid-soluble mercury is below the limit.

Unless otherwise noted, all concentrations reported herein are expected to be essentially equivalent to acid-soluble mercury concentrations. All concentrations are expressed as mercury, not as the chemical tested. The criteria presented herein supersede previous aquatic life water quality criteria for mercury (U.S. EPA, 1976, 1980) because these new criteria were derived using improved procedures and additional information. Whenever adequately justified, a national criterion may be replaced by a site-specific criterion (U.S. EPA, 1983b), which may include not only site-specific criterion concentrations (U.S. EPA, 1983c), but also site-specific durations of averaging periods and site-specific frequencies of allowed exceedences (U.S. EPA, 1985). The latest literature search for information for this document was conducted in May, 1984; some newer information was also used.

Acute Toxicity to Aquatic Animals

Table 1 contains the primary acute toxicity data for three classes of mercury compounds: mercury(II), methylmercury, and other mercury compounds, chiefly organic. The latter information exists principally because many of these compounds were considered for use in treatment of diseases and control of parasites in fish culture, although their source for environmental concern is from industrial and agricultural uses for fungus control. Both phenylmercuric acetate and pyridylmercuric acetate have been called PMA. Tests have been conducted on different formulations which contain various percentages of active ingredient and the percentages of active ingredient given by the authors were used to calculate mercury concentrations. When the percentage of active ingredient was not given for pyridylmercuric acetate, 80 percent was assumed (Allison, 1957).

The freshwater acute toxicity values indicate that the difference in sensitivity between different species to a particular mercury compound is far greater than the difference in sensitivity of a particular species to various mercury compounds. For example, the reported acute values for mercury(II) range from 2.217 µg/L for Daphnia pulex to 2,000 µg/L for the aquatic stages of certain insects, with a continual gradation in sensitivity among intermediate species (Table 3). On the other hand, Joshi and Rege (1980), Lock and van Overbeeke (1981) and Matida, et al. (1971) found that various species were 4 to 31 times more sensitive to various organic mercury compounds than to mercuric chloride (Table 1).

MacLeod and Pessah (1973) studied the effect of temperature on the acute toxicity of mercuric chloride to rainbow trout. At 5, 10, and 15 C, the LC50s were 400, 280, and 220 ug/L, respectively (Tables 1 and 6). Clemens and Sneed (1958b) found a similar effect of temperature on toxicity to

juvenile channel catfish; at 10, 16.5, and 24 C the acute values for phenylmercuric acetate were 1,960, 1,360, and 233 µg/L, respectively (Table 6).

The 28 Genus Mean Acute Values in Table 3 were calculated as geometric means of the available Species Mean Acute Values (Tables 1 and 3). Acute values are available for more than one species in each of two genera, and the range of Species Mean Acute Values within each genus is less than a factor of 1.6. On the other hand, a midge was among the most sensitive species, whereas other insects were the most resistant species. The most sensitive genus, Daphnia, is 756 times more sensitive than the most resistant, Acroneuria (Table 3). A freshwater Final Acute Value of 4.857 µg/L was obtained for mercury(II) using the Genus Mean Acute Values in Table 3 and the calculation procedure described in the Guidelines. Not enough data are available to calculate a Final Acute Value for methylmercury, but the available data indicate that it is more acutely toxic than mercury(II).

Saltwater fishes and invertebrates both show wide ranges of sensitivities to mercury(II). Acute values for fishes range from 36 µg/L for spot to 1,678 µg/L for winter flounder (Tables 1 and 3). Among invertebrates a mysid has an acute value of 3.5 µg/L, whereas the value for the soft-shell clam is 400 µg/L. Of the 29 saltwater genera for which acute values are available, the most sensitive, Mysidopsis, is 479 times more sensitive than the most resistant, Pseudopleuronectes. Acute values are available for more than one species in each of three genera and the range of Species Mean Acute Values within each genus is less than a factor of 1.7. The saltwater Final Acute Value of 4.125 µg/L was calculated for mercury(II) from the Genus Mean Acute Values in Table 3.

Chronic Toxicity to Aquatic Animals

Chronic toxicity tests with Daphnia magna have been conducted on three mercury compounds (Table 2). The renewal and flow-through techniques produced similar results for mercury(II), but the renewal technique produced much higher results for methylmercury, presumably because of volatility. In addition, a chronic test with brook trout on methylmercuric chloride yielded a chronic value of 0.5193 µg/L. Both an early life-stage test and a life-cycle test on mercuric chloride found adverse effects on the fathead minnow at all concentrations tested including the lowest of 0.23 µg/L. For mercuric chloride the acute-chronic ratio with Daphnia magna is less than 6, whereas that with the fathead minnow is greater than 600. For methylmercury the acute-chronic ratio with brook trout is 142.3.

A chronic value of 1.131 µg/L was obtained in a flow-through life-cycle exposure of a mysid to mercuric chloride (Table 2). Groups of 30 juvenile mysids were reared in each of 5 concentrations for 36 days at 21 C and a salinity of 30 g/kg. Effects examined included time to first spawn and productivity (total number of young/number of available female spawning days and total number of spawns/number of available female spawning days). No spawning occurred at 2.5 µg/L. Time to spawn and productivity at 1.6 µg/L were significantly different from the controls. The highest concentration at which no statistically significant effect on reproductive processes was detected was 0.8 µg/L. Therefore, the chronic limits are 0.8 and 1.6 µg/L and the chronic value is 1.131 µg/L. The 96-hr LC50 for this species in the same study was 3.5 ug/L, giving an acute-chronic ratio of 3.095 (Table 2).

The species mean acute-chronic ratio for Daphnia magna is 4.498, whereas that for the mysid is 3.095 (Table 3). These are sensitive species in fresh and salt water, respectively, and the four most sensitive species in each

water are invertebrates. Thus, it seems reasonable to use the geometric mean of these two values as the Final Acute-Chronic Ratio (Table 3). Division of the Final Acute Values by 3.731 results in freshwater and saltwater Final Chronic Values of 1.302 and 1.106 µg/L, respectively. Even though the fathead minnow was considerably less sensitive than Daphnia magna in acute tests, the acute-chronic ratio for the fathead minnow is so high that its chronic value is below the Final Chronic Value and probably below the Final Residue Value (see below). If the acute-chronic ratio of greater than 649 for the fathead minnow is representative of ratios for other freshwater and saltwater fishes, then twelve of fourteen tested fish species, including the rainbow trout, coho salmon, bluegill, and haddock, would have chronic values below the Final Chronic Value. Various values for vertebrates in Table 6 are below the Final Chronic Value or are indicative of large acute-chronic ratios.

Toxicity to Aquatic Plants

Whereas some freshwater plant values for mercury(II) are about 1,000 µg/L (Table 4), effects of mercury(II) and methylmercury have been observed at concentrations below 10 µg/L, respectively (Table 6). Some organomercury compounds have affected algae at concentrations less than 1.0 µg/L (Table 6). Although freshwater plants are relatively insensitive to mercury(II) and sensitive to methylmercury, they do not appear to be more sensitive to methylmercury than freshwater animals.

Data concerning the toxicity of mercuric chloride to saltwater plants are from four studies with eight species of algae. The EC50s (Table 4) indicate reduction in growth at concentrations ranging from 10 to 160 µg/L. No data are available concerning the toxicities of organomercury compounds to saltwater plants.

Bioaccumulation

Bioconcentration is a function of the relative rates of uptake and depuration. The bioconcentration factor (BCF) of mercury is high for fish because uptake is relatively fast and depuration is very slow. Thus, the biological half-life of mercury in fish is approximately 2 to 3 years (de Freitas, et al. 1974, 1977; Jarvenpaa, et al. 1970; Lockhart, et al. 1972; McKim, et al. 1976; Mellinger, 1973; Ruohtula and Miettinen, 1975; Sharpe, et al. 1977). Depuration of mercury is so slow that, even in the absence of exposure to mercury, long-term reduction in the concentration of mercury in fish tissue is largely due to dilution by tissue addition from growth. Usually less than 60 percent of mercury in invertebrates is methylated, but in fish, except for young fish, usually more than 70 percent is methylated (Bache, et al. 1971; Baluja, et al. 1983; Busch, 1983; Cappon, 1984; Cappon and Smith, 1982a,b; Hattula, et al. 1978; Hildebrand, et al. 1980; Huckabee, et al. 1979; Kudo, et al. 1982; Luten, et al. 1980; Paasivirta, et al. 1983).

The distribution of mercury within a fish is the result of the movement of mercury from the absorbing surfaces (gills, skin, and gastrointestinal tract), into the blood, then to the internal organs, and eventually either to the kidney or bile for recycling or elimination or to muscle for long-term storage. As the tissue concentration approaches steady-state, net accumulation rate is slowed either by a reduction in uptake rate, possibly due to inhibition of membrane transport, or by an increase in depuration rate, possibly because of a saturation of storage sites, or both.

High concentrations of mercury in the slime coat of certain freshwater fishes, such as burbot, eels, and northern pike, and in the skin of acutely-exposed fishes are believed to be due to the methylating activity of bacteria

prevalent in the mucous coat (Jernelov, 1968). In addition, acutely toxic concentrations of mercury have been reported to stimulate secretion of mucus (Baker, 1973; Lock, et al. 1981; McKone, et al. 1971). When acutely exposed fish are placed in mercury-free water, the skin quickly loses mercury, probably because of elimination of mercury(II) and sloughing of the slime coat (Burrows and Krenkel, 1973; Burrows, et al. 1974). The skin and mucous coat are in direct contact with mercury in water and can accumulate proportionately more mercury during short exposure than muscle. During long exposure there is sufficient time for mercury to reach more permanent storage sites.

Because sorption at the gill surface is a major pathway of mercury into aquatic organisms (Fromm, 1977), increases in temperature and activity cause increases in metabolic rate and ventilation rate and, therefore, uptake rate (de Freitas and Hart, 1975; Rogers and Beamish, 1981). The relationship between temperature and tissue residues seems to apply primarily before steady-state is reached (Reinert, et al. 1974) but also to some extent at steady-state (Boudou, et al. 1979; Cember, et al. 1978; Hartung, 1976). The latter is difficult to understand if steady-state results from saturation of available binding sites. Apparently not only are uptake and depuration accelerated by temperature (Ruohtula and Miettinen, 1975), but higher tissue residues also occur at higher temperatures, posibly because the uptake rate increases proportionately more than the depuration rate.

Similarly, low concentrations of dissolved oxygen are likely to increase both respiration rate and uptake rate. Larson (1976) found that the low concentration of dissolved oxygen in a eutrophic lake forced fishes into warmer surface waters to secure adequate oxygen. The warmer surface water apparently stimulated metabolic rate and increased mercury uptake.

Increased metabolic rate increases not only ventilation rate but also energy demand and thus increases food consumption and exposure to mercury through the food chain. Uptake through both the gills and the digestive tract is significant for fish, and some data suggest that tissue residues are higher in organisms exposed via both routes than via either separately (Boudou, et al. 1979; Phillips and Buhler, 1978). The relative importance of uptake from food for various fish species depends on such things as assimilation efficiency (Phillips and Gregory, 1979), body size, growth rate, and life span (Sharpe, et al. 1977), and diet (Murray, 1978). Although Murray (1978) did find different concentrations of mercury in different fish species, Huckabee (1972) and Huckabee, et al. (1974) found similar concentrations in both forage and game fish in the same environment.

Haines (1981) reported that acid rain tends to scour more mercury from the air. Acidification of a body of water might also increase mercury residues in fish even if no new input of mercury occurs, possibly because lower pH increases ventilation rate and membrane permeability, accelerates the rates of methylation and uptake, affects partitioning between sediment and water, or reduces growth or reproduction of fish (Akielaszek and Haines, 1981; Fromm, 1980; Hahne and Kroontje, 1973; Jernelov, 1980; Miller and Akagi, 1979; Ribeyre and Boudou, 1982; Rudd and Turner, 1983b; Scheider, et al. 1979; Stokes, et al. 1983; Tsai, et al. 1975; Wrenn and MacCrimmon, 1983). However, Heiskary and Helwig (1983) did not find a relation between pH and mercury in fish.

The available information (e.g., Boudou and Ribeyre, 1981; Boudou, et al. 1977, 1980; de Freitas, et al. 1981; Hamdy and Prabhu, 1979; Hamelink, et al. 1977; Herrick, et al. 1982; Huckabee, et al. 1979; Jernelov and Lann, 1971; Klaverkamp, et al. 1983c; MacCrimmon, et al. 1983; Norstrom, et al.

1976; Phillips, et al. 1980; Ribeyre, et al. 1980; Rogers and Beamish, 1982; Rogers and Qadri, 1982) indicates that the importance of uptake from food probably depends on the form and concentration of mercury in the diet and on the size and metabolic rate of the fish. Transfer of mercury from fish to wildlife predators has also been observed (Heinz, et al. 1980; Kucera, 1983; Wren, et al. 1983).

The available freshwater BCFs are listed in Tables 5 and 6. Table 5 contains BCFs only from those studies in which the exposure concentrations were adequately measured and the tissue residues reached steady-state or the test lasted longer than 27 days. Although the BCFs presented in Table 6 do not meet all these conditions, they do provide information on BCFs for plants and illustrate the very important influence of temperature on bioconcentration.

McKim, et al. (1976) studied the uptake of methylmercury into various tissues of brook trout. At concentrations in water of 0.93, 0.29, 0.09 and 0.03 µg/L the resulting concentrations in muscle after 273 days were 10,000, 5,000, 1,900, and 1,000 µg/kg and the corresponding BCFs were 11,000, 17,000, 21,000, and 33,000, respectively. Because the concentration of mercury in the muscle did not decrease as much as the concentration in water, the BCFs increased as the concentration in water decreased. A possible explanation for an inverse relationship between concentration in water and BCF is that steady-state results from saturation of available binding sites (Cember, et al. 1978). The maximum concentration in tissue would then be dependent on the number of available binding sites and would be independent of the concentration of mercury in water. If the concentration in tissue were constant, the BCF would be inversely proportional to the concentration in water. However, neither the concentration in tissue nor the BCF was constant. The

comparable BCFs for the whole body were 10,000, 12,000, 12,000, and 23,000 and are lower than those for muscle. The fish were adversely affected at 0.93 µg/L, but for both muscle and whole body, the BCF at this concentration is in line with the other BCFs. Even though concentrations up to 0.29 µg/L did not cause statistically significant adverse effects, the concentration of mercury in fish exposed to 0.03 µg/L were at the FDA action level.

Olson, et al. (1975) obtained much higher BCFs for methylmercury with the fathead minnow, and the BCF was also concentration-dependent. As the concentration in the water decreased from 0.247 µg/L to 0.018 µg/L, the concentration in the fish decreased from 10,900 µg/kg to 1,470 µg/kg, but the BCF decreased from 44,100 to 81,700. The contrast between the results with the fathead minnow at 25 C (Olson, et al. 1975) and brook trout at 9 to 15 C (McKim, et al. 1976) is one of considerable interest and potential importance. The trout were fed pelleted feed, and so had little opportunity for food chain input. In contrast, the fathead minnow is a browser and probably fed not only on the introduced food but also on the Aufwuchs growing in the test solution to which mercury had been added. Thus the higher BCFs for the fathead minnow might be more representative of field situations in which fish are exposed to methylmercury via both the water and food (Phillips and Buhler, 1978; Phillips and Gregory, 1979; Rogers and Beamish, 1982). Also, because temperature affects bioconcentration, the fathead minnow might be more representative of commonly consumed warmwater fishes.

In a 75-day test, Niimi and Lowe-Jinde (1984) found 12 mg/kg in the whole body of rainbow trout exposed to 0.15 µg methylmercury/L (0.14 µg mercury/L) in water and 18 µg mercury/kg in food. The BCF of 85,700 is higher than the highest BCF obtained by Olson, et al. (1975). However, at 0.012 µg/L and 18 ug/kg, they found 0.053 mg/kg in the trout. This BCF of

4,077 is lower than the lowest BCF obtained by McKim, et al. (1976). Also, although both McKim, et al. (1976) and Olson, et al. (1975) found higher BCFs at lower concentrations in water, Niimi and Lowe-Jinde (1984) found just the opposite.

The FDA action level for mercury in fish and shellfish is 1.0 mg/kg (Table 5), and now only applies to the methylmercury in consumed tissues (U.S. FDA, 1984a,b). In their test on methylmercury, McKim, et al. (1976) found that brook trout exposed to 0.03 ug/L contained 1 mg/kg in muscle tissue. However, in their test on methylmercury with the fathead minnow, Olson, et al. (1975) found that exposure to 0.018 µg/L resulted in 1.47 mg/kg in the fish and a BCF of 81,700. Use of this BCF with the FDA action level results in a Final Residue Value of 0.012 µg/L for methylmercury (Table 5). The concentration in the fathead minnow is for whole body, but Heisinger, et al. (1979) found no significant difference between various body compartments. Further, Huckabee, et al. (1974) found that all fishes in a particular environment acquired about the same concentrations of mercury in both the whole body and muscle tissue when they were chronically exposed to low concentrations of mercury. On the other hand, Heiskary and Helwig (1983) and McKim, et al. (1976) found higher concentrations of mercury in the edible portion of fish than in the whole body. Thus the concentration of mercury in the muscle of some edible species is likely to exceed the FDA action level when exposed to methylmercury at a concentration of 0.012 µg/L.

Although the FDA action level only applies to methylmercury in fish and shellfish, it can be used to derive a water quality criterion for mercury(II) because most of the mercury in fish is methylmercury even if the organisms were exposed to inorganic mercury (de Freitas, et al. 1974; Jernelov and Lann, 1971). A BCF of 4,994 was obtained for mercuric chloride in a

life-cycle test with the fathead minnow (Snarski and Olson, 1982). This BCF is based on the concentration of mercuric chloride in the water and the total concentration of organic and inorganic mercury in the tissue. Even though all concentrations tested caused adverse effects and the higher concentrations caused more severe effects, the BCFs were similar at all concentrations and were lower than those obtained with methylmercury by McKim, et al. (1976) and Olson, et al. (1975). Use of the BCF of 4,994 with the FDA action level of 1.0 mg/kg results in a freshwater Final Residue Value of 0.20 µg/L for mercury(II) (Table 5). This value of 0.20 µg/L derived for mercury(II) would, however, be too low if field BCFs are higher than laboratory BCFs, if waters contain substantial concentrations of methylmercury, or if methylation processes are accelerated in sediment.

Information on the bioconcentration of various mercury compounds by saltwater animals and plants is included in Tables 5 and 6 and by saltwater plankton in Table 6. For mercuric chloride, BCFs ranged from 853 to 10,920 with algae. In tests with the eastern oyster, BCFs of 10,000, 40,000, and 40,000 were obtained for mercuric chloride, methylmercuric chloride and phenylmercuric chloride, respectively (Kopfler, 1974). These are similar to the BCFs obtained with freshwater fish, but the BCF of 129 obtained for mercuric chloride in tail muscle of the American lobster is much lower.

To protect the marketability of saltwater shellfish for human consumption, Final Residue Values can be calculated based on the BCFs for the oyster and the FDA action level of 1.0 mg/kg. Accordingly, the Final Residue Values for mercuric chloride, methylmercuric chloride, and phenylmercuric chloride are 0.10, 0.025, and 0.025 µg/L, respectively. However, at these concentrations fifty percent of the exposed oysters would probably exceed the

FDA action level if all the mercury in the body were present as methylmercury.

Other Data

Most of the significant freshwater and saltwater results in Table 6 have already been discussed in connection with data in Tables 1-5, but a few additional items deserve special mention. Comparable tests with four species showed that mercuric cyanide was 0.67 to 50 times as toxic as mercuric chloride. Also, Birge, et al. (1979) reported that flow-through tests gave EC50s nearly two orders of magnitude lower than static tests with rainbow trout, catfish, goldfish, and largemouth bass (Table 6). Bouquegneau (1979) found that preexposure induced metallothionein production, which then protected the fish.

Molybdenum (Yamane and Koizumi, 1982) and vitamin E (Ganther, 1978, 1980) affects the toxicity of mercury to mammals, and probably many consumers of aquatic organisms, as does selenium (e.g., Alexander, et al. 1979; Berlin, 1978; National Research Council, 1978; National Research Council Canada, 1979; Stopford and Goldwater, 1975; Strom, et al. 1979). Wobeser, et al. (1976a,b) found methylmercury to be much less toxic to mink when they were fed freshwater drum, Aplodinolus grunniens, containing high mercury tissue residues than when they were fed a diet to which methylmercury chloride had been added. On the other hand, Albanus, et al. (1972) and Charbonneau, et al. (1974) found similar toxicity to cats when fed similar dietary concentrations of methylmercury, one as a tissue residue in pike and the other with methylmercury added to the ration.

Finley and Stendell (1978) and Heinz (1979a) fed black and mallard ducks, respectively, food contaminated with methylmercuric dicyandiamide.

These feeding studies extended over two and three generations, respectively, and demonstrated reduced hatching success and juvenile survival at mercury concentrations that were estimated to be equivalent to 0.5 and 0.1 mg/kg, respectively, in the natural succulent food of the wild ducks. These results were not used to estimate a Final Residue Value based on food for wildlife because the dicyandiamide compound might not represent the toxicity of methylmercury alone. Nevertheless, these tests suggest that the Final Residue Value might be an order of magnitude too high because at least one of these authors believes that the anion had little effect on the results (Heinz, 1979b).

Unused Data

Some data on the effects of mercury on aquatic organisms were not used because the studies were conducted with species that are not resident in North America (e.g., Ahsanullah, 1982; Akiyama, 1970; Dial, 1978a,b; Heisinger and Green, 1975; Jones, 1939a, 1940, 1973, 1975; Khangarot, et al. 1982; Kihlstrom and Hulth, 1972; Krishnaja and Rege, 1982; Mathur, et al. 1981; McClurg, 1984; Murti and Shukla, 1984; Nagashima, et al. 1983; Saxena and Parashari, 1983; Shaffi, 1981; Srivastava, 1982; van den Broek and Tracey, 1981; Verma, et al. 1984), or because the test species was not obtained in North America and was not identified well enough to determine if it is resident in North America (Hannerz, 1968). Brown and Ahsanullah (1971) conducted tests with brine shrimp, which species is too atypical to be used in deriving national criteria. Reviews by Chapman, et al. (1968), Eisler (1981), Eisler, et al. (1979), Phillips and Russo (1978), and Thompson, et al. (1972) only contain data that have been published elsewhere.

The 96-hr values reported by Buikema, et al. (1974a,b) were subject to error because of possible reproductive interactions (Buikema, et al. 1977). Applegate, et al. (1957) exposed only one or two organisms. Data were not used if the mode of exposure was inappropriate for deriving water quality criteria (Giblin and Massaro, 1973; Lucu and Skreblin, 1981; Schmidt-Nielson, et al. 1977; Weisbart, 1973). In addition, data were not used if mercury was a component of an effluent (Wong, et al. 1982) or if the test organisms were cultured in one water and tested in another (Bringmann and Kuhn, 1982). Bills, et al. (1977) and Passino and Cotant (1979) did not report any usable results. Jones (1935, 1938, 1939b, 1947), Miller (1980, 1981), and Nuzzi (1972) did not report a clearly defined endpoint.

Results of some laboratory tests were not used because the test was conducted in distilled water (McKone, et al. 1971), the quality of the dilution water or medium was questionable (Brkovic-Popovic and Popovic, 1977a,b; Carter and Cameron, 1973; Kim, et al. 1977a,b; Stary and Kratzer, 1982; Stary, et al. 1982, 1983), or because the test solution or culture medium contained too much EDTA which would probably complex mercury (Gutierrez-Galindo, 1981; Knowles and Zingmark, 1978; Stratton and Corke, 1979; Stratton, et al. 1979).

Data concerning concentrations of mercury in wild organisms (e.g., Abernathy and Cumbie, 1977; Armstrong and Scott, 1979; Bodaly, et al. 1984; Copeland and Ayers, 1972; DiNardi, et al. 1974; Flegal, et al. 1981; Helwig and Hora, 1983; Hildebrand, et al. 1980; Jensen, et al. 1981; Leonzio, et al. 1982; Martin, et al. 1984; May and McKinney, 1981; Mitchell, et al. 1982; Moore and Sutherland, 1980; Murray, 1978; Pennington, et al. 1982; Phillips and Buhler, 1980; Price and Knight, 1978; Ray, et al. 1984; Sheffy, 1978; Tsui and McCart, 1981; Wachs, 1982; Watling, et al. 1981) were not used to

calculate bioaccumulation factors if the concentrations of mercury in the ambient water during the period of exposure was not adequately measured. Studies using isotopic mercury (e.g., Cunningham and Tripp, 1975; Glooschenko, 1969) were not used because of the possibility of isotopic discrimination.

Results of bioconcentration tests were not used if the tests were conducted in distilled water, were not long enough, were not flow-through, or if the concentration of mercury in the test solution was not adequately measured (e.g., Cunningham and Tripp, 1973; Kim, et al. 1977a,b; McKone, et al. 1971; Medeiros, et al. 1980; Middaugh and Rose, 1974; Phillips and Gregory, 1980; Ribeyre, et al. 1980; Sharma and Davis, 1980a; Scary and Kratzer, 1980, 1982; Scary, et al. 1980, 1981, 1982, 1983; Vernberg and O'Hara, 1972).

Summary

Data are available on the acute toxicity of mercury(II) to 28 genera of freshwater animals. Acute values for invertebrate species range from 2.2 µg/L for *Daphnia pulex* to 2,000 µg/L for three insects. Acute values for fishes range from 30 µg/L for the guppy to 1,000 µg/L for the Mozambique tilapia. Few data are available for various organomercury compounds and mercurous nitrate, and they all appear to be 4 to 31 times more acutely toxic than mercury(II).

Available chronic data indicate that methylmercury is the most chronically toxic of the tested mercury compounds. Tests on methylmercury with *Daphnia magna* and brook trout produced chronic values less than 0.07 ug/L. For mercury(II) the chronic value obtained with *Daphnia magna* was about 1.1 µg/L and the acute-chronic ratio was 4.5. In both a life-cycle

test and an early life-stage test on mercuric chloride with the fathead minnow, the chronic value was less than 0.26 µg/L and the acute-chronic ratio was over 600.

Freshwater plants show a wide range of sensitivities to mercury, but the most sensitive plants appear to be less sensitive than the most sensitive freshwater animals to both mercury(II) and methylmercury. A bioconcentration factor of 4,994 is available for mercury(II), but the bioconcentration factors for methylmercury range from 4,000 to 85,000.

Data on the acute toxicity of mercuric chloride are available for 29 genera of saltwater animals including annelids, molluscs, crustaceans, echinoderms, and fishes. Acute values range from 3.5 µg/L for a mysid to 1,678 µg/L for winter flounder. Fishes tend to be more resistant and molluscs and crustaceans tend to be more sensitive to the acute toxic effects of mercury(II). Results of a life-cycle test with the mysid show that mercury(II) at a concentration of 1.6 µg/L significantly affected time of first spawn and productivity; the resulting acute-chronic ratio was 3.1.

Concentrations of mercury that affected growth and photosynthetic activity of one saltwater diatom and six species of brown algae range from 10 to 160 µg/L. Bioconcentration factors of 10,000 and 40,000 have been obtained for mercuric chloride and methylmercury with an oyster.

National Criteria

Derivation of a water quality criterion for mercury is more complex than for most metals because of methylation of mercury in sediment, in fish, and in the food chain of fish. Apparently almost all mercury currently being discharged is mercury(II). Thus mercury(II) should be the only important

possible cause of acute toxicity and the Criterion Maximum Concentrations can be based on the acute values for mercury(II).

The best available data concerning long-term exposure of fish to mercury(II) indicates that concentrations above 0.23 µg/L caused statistically significant effects on the fathead minnow and caused the concentration of total mercury in the whole body to exceed 1.0 mg/kg. Although it is not known what percent of the mercury in the fish was methylmercury, it is also not known whether uptake from food would increase the concentration in the fish in natural situations. Species such as rainbow trout, coho salmon, and especially the bluegill, might suffer chronic effects and accumulate high residues of mercury about the same as the fathead minnow.

With regard to long-term exposure to methylmercury, McKim, et al. (1976) found that brook trout can exceed the FDA action level without suffering statistically significant adverse effects on survival, growth, or reproduction. Thus for methylmercury the Final Residue Value would be substantially lower than the Final Chronic Value.

Basing a freshwater criterion on the Final Residue Value of 0.012 µg/L derived from the bioconcentration factor of 81,700 for methylmercury with the fathead minnow (Olson, et al. 1975) essentially assumes that all discharged mercury is methylmercury. On the other hand, there is the possibility that in field situations uptake from food might add to the uptake from water. Similar considerations apply to the derivation of the saltwater criterion of 0.025 µg/L using the BCF of 40,000 obtained for methylmercury with the Eastern oyster (Kopfler, 1974). Because the Final Residue Values for methylmercury are substantially below the Final Chronic Values for mercury(II), it is probably not too important that many fishes, including the rainbow trout,

coho salmon, bluegill, and haddock might not be adequately protected by the freshwater and saltwater Final Chronic Values for mercury(II).

In contrast to all the complexities of deriving numerical criteria for mercury, monitoring for unacceptable environmental effects should be relatively straightforward. The most sensitive adverse effect will probably be exceedence of the FDA action level. Therefore, existing discharges should be acceptable if the concentration of methylmercury in the edible portion of exposed consumed species does not exceed the FDA action level.

The procedures described in the "Guidelines for Deriving Numerical National Water Quality Criteria for the Protection of Aquatic Organisms and Their Uses" indicate that, except possibly where a locally important species is very sensitive, freshwater aquatic organisms and their uses should not be affected unacceptably if the four-day average concentration of mercury does not exceed 0.012 µg/L more than once every three years on the average and if the one-hour average concentration does not exceed 2.4 µg/L more than once every three years on the average. If the four-day average concentration exceeds 0.012 µg/L more than once in a three-year period, the edible portion of consumed species should be analyzed to determine whether the concentration of methylmercury exceeds the FDA action level.

The procedures described in the "Guidelines for Deriving Numerical National Water Quality Criteria for the Protection of Aquatic Organisms and Their Uses" indicate that, except possibly where a locally important species is very sensitive, saltwater aquatic organisms and their uses should not be affected unacceptably if the four-day average concentration of mercury does not exceed 0.025 µg/L more than once every three years on the average and if the one-hour average concentration does not exceed 2.1 µg/L more than once every three years on the average. If the four-day average concentration

exceeds 0.025 µg/L more than once in a three-year period, the edible portion of consumed species should be analyzed to determine whether the concentration of methylmercury exceeds the FDA action level.

EPA believes that a measurement such as "acid-soluble" would provide a more scientifically correct basis upon which to establish criteria for metals. The criteria were developed on this basis. However, at this time, no EPA approved methods for such a measurement are available to implement the criteria through the regulatory programs of the Agency and the States. The Agency is considering development and approval of methods for a measurement such as "acid-soluble". Until available, however, EPA recommends applying the criteria using the total recoverable method. This has two impacts: (1) certain species of some metals cannot be analyzed directly because the total recoverable method does not distinguish between individual oxidation states, and (2) these criteria may be overly protective when based on the total recoverable method.

The recommended exceedence frequency of three years is the Agency's best scientific judgment of the average amount of time it will take an unstressed system to recover from a pollution event in which exposure to mercury exceeds the criterion. Stressed systems, for example, one in which several outfalls occur in a limited area, would be expected to require more time for recovery. The resilience of ecosystems and their ability to recover differ greatly, however, and site-specific criteria may be established if adequate justification is provided.

The use of criteria in designing waste treatment facilities requires the selection of an appropriate wasteload allocation model. Dynamic models are preferred for the application of these criteria. Limited data or other factors may make their use impractical, in which case one should rely on a

steady-state model. The Agency recommends the interim use of 1Q5 or 1Q10 for Criterion Maximum Concentration (CMC) design flow and 7Q5 or 7Q10 for the Criterion Continuous Concentration (CCC) design flow in steady-state models for unstressed and stressed systems respectively. These matters are discussed in more detail in the Technical Support Document for Water Quality-Based Toxics Control (U.S. EPA, 1985).

Table 1. Acute Toxicity of Mercury to Aquatic Animals

Species	Method*	Chemical	LC50 or EC50 (µg/L)**	Species Mean Acute Value (µg/L)**	Reference
		FRESHWATER SPECIES			
		Mercury(II)			
Tubificid worm, Branchiura sowerbyi	R, U	Mercuric chloride	80	80	Chapman, et al. 1982a
Tubificid worm, Limnodrilus hoffmeisteri	R, U	Mercuric chloride	180	180	Chapman, et al. 1982a,b
Tubificid worm, Quistadrilus multisetosus	R, U	Mercuric chloride	250	250	Chapman, et al. 1982a
Tubificid worm, Rhyacodrilus montana	R, U	Mercuric chloride	240	240	Chapman, et al. 1982a
Tubificid worm, Spirosperma ferox	R, U	Mercuric chloride	330	330	Chapman, et al. 1982a
Tubificid worm, Spirosperma nikolskyi	R, U	Mercuric chloride	500	500	Chapman, et al. 1982a
Tubificid worm, Stylodrilus heringianus	R, U	Mercuric chloride	140	140	Chapman, et al. 1982a
Tubificid worm, Tubifex tubifex	R, U	Mercuric chloride	140	140	Chapman, et al. 1982a,b
Tubificid worm, Varichaeta pacifica	R, U	Mercuric chloride	100	100	Chapman, et al. 1982a
Worm, Nais sp.	S, M	Mercuric nitrate	1,000	1,000	Rehwoldt, et al. 1973
Snail (embryo), Amnicola sp.	S, M	Mercuric nitrate	2,100***	-	Rehwoldt, et al. 1973
Snail (adult), Amnicola sp.	S, M	Mercuric nitrate	80	80	Rehwoldt, et al. 1973
Snail, Aplexa hypnorum	S, U	Mercuric chloride	370	370	Holcombe, et al. 1983

Table 1. (Continued)

Species	Method[a]	Chemical	LC50 or EC50 (µg/L)**	Species Mean Acute Value (µg/L)**	Reference
Cladoceran, *Daphnia magna*	S, U	Mercuric chloride	<4.4****	-	Anderson, 1948
Cladoceran, *Daphnia magna*	S, U	Mercuric chloride	5	-	Biesinger & Christensen, 1972
Cladoceran, *Daphnia magna*	S, U	Mercuric chloride	3.177	-	Canton & Adema, 1978
Cladoceran, *Daphnia magna*	S, U	Mercuric chloride	1.478	-	Canton & Adema, 1978
Cladoceran, *Daphnia magna*	S, U	Mercuric chloride	2.180	-	Canton & Adema, 1978
Cladoceran (<6 hr old), *Daphnia magna*	S, U	Mercuric chloride	4.4	-	Barera & Adams, 1983
Cladoceran (<24 hr old), *Daphnia magna*	S, U	Mercuric chloride	4.4	-	Barera & Adams, 1983
Cladoceran (1-9 day old), *Daphnia magna*	S, U	Mercuric chloride	5.2-14.8***	3.157	Barera & Adams, 1983
Cladoceran, *Daphnia pulex*	S, U	Mercuric chloride	2.217	2.217	Canton & Adema, 1978
Amphipod, *Gammarus* sp.	S, M	Mercuric nitrate	10	10	Rehwoldt, et al. 1973
Crayfish (male, mixed ages), *Faxonella clypeatus*	R, M	Mercuric chloride	20	20	Holt & Fingerman, 1977; Holt, 1981
Crayfish, *Orconectes limosus*	S, M	Mercuric chloride	50	50	Boutet & Chaisemartin, 1973
Mayfly, *Ephemerella subvaria*	S, U	Mercuric chloride	2,000	2,000	Warnick & Bell, 1969
Damselfly, (Unidentified)	S, M	Mercuric nitrate	1,200	1,200	Rehwoldt, et al. 1973

Table 1. (Continued)

Species	Method[a]	Chemical	LC50 or EC50 (µg/L)[aa]	Species Mean Acute Value (µg/L)[aa]	Reference
Stonefly, *Acroneuria lycorias*	S, U	Mercuric chloride	2,000	2,000	Warnick & Bell, 1969
Caddisfly, *Hydropsyche bettani*	S, U	Mercuric chloride	2,000	2,000	Warnick & Bell, 1969
Caddisfly, (Unidentified)	S, M	Mercuric nitrate	1,200	1,200	Rehwoldt, et al. 1973
Midge, *Chironomus* sp.	S, M	Mercuric nitrate	20	20	Rehwoldt, et al. 1973
Coho salmon (juvenile), *Oncorhynchus kisutch*	R, M	Mercuric chloride	240	240	Lorz, et al. 1978
Rainbow trout (juvenile), *Salmo gairdneri*	R, U	Mercuric chloride	155.1	-	Matida, et al., 1971
Rainbow trout (juvenile), *Salmo gairdneri*	FT, U	Mercuric chloride	280	-	MacLeod & Pessah, 1973
Rainbow trout (juvenile), *Salmo gairdneri*	FT, U	Mercuric chloride	220	-	MacLeod & Pessah, 1973
Rainbow trout, *Salmo gairdneri*	FT, U	Mercuric chloride	420	-	Daoust, 1981
Rainbow trout (juvenile), *Salmo gairdneri*	FT, M	Mercuric chloride	275	275	Lock & van Overbeeke, 1981
Fathead minnow, *Pimephales promelas*	FT, M	Mercuric chloride	168	-	Snarski & Olson, 1982
Fathead minnow, *Pimephales promelas*	FT, M	Mercuric chloride	150	158.7	Call, et al. 1983
Mosquitofish (female), *Gambusia affinis*	S, U	Mercuric chloride	180	180	Joshi & Rege, 1980
Guppy (116-157 mg), *Poecilia reticulata*	R, U	Mercuric chloride	30	-	Deshmukh & Marathe, 1980

Table 1. (Continued)

Species	Method[a]	Chemical	LC50 or EC50 (µg/L)[**]	Species Mean Acute Value (µg/L)[**]	Reference
Guppy (362-621 mg), *Poecilia reticulata*	R, U	Mercuric chloride	53.5[***]	30	Deshmukh & Marathe, 1980
Bluegill (juvenile), *Lepomis macrochirus*	S, U	Mercuric chloride	160	160	Holcombe, et al. 1983
Mozambique tilapia, *Tilapia mossambica*	S, U	Mercuric chloride	1,000	1,000	Qureshi & Saksena, 1980
		Methylmercury			
Rainbow trout (juvenile), *Salmo gairdneri*	R, U	Methylmercuric chloride	25	-	Matida, et al. 1971
Rainbow trout (larva), *Salmo gairdneri*	R, U	Methylmercuric chloride	24	-	Wobeser, 1973
Rainbow trout (juvenile), *Salmo gairdneri*	R, U	Methylmercuric chloride	42	-	Wobeser, 1973
Rainbow trout (juvenile), *Salmo gairdneri*	FT, M	Methylmercuric chloride	24	24	Lock & van Overbeeke, 1981; Lock, et al. 1981
Brook trout (juvenile), *Salvelinus fontinalis*	FT, M	Methylmercuric chloride	84	-	McKim, et al. 1976
Brook trout (yearling), *Salvelinus fontinalis*	FT, M	Methylmercuric chloride	65	73.89	McKim, et al. 1976
		Other Mercury Compounds			
Rainbow trout (juvenile), *Salmo gairdneri*	R, U	Phenylmercuric acetate	5	5	Matida, et al. 1971
Rainbow trout (2 mos), *Salmo gairdneri*	FT, M	Mercurous nitrate	33.0	33.0	Hale, 1977
Goldfish, *Carassius auratus*	S, U	Phenylmercuric lactate	82	82	Ellis, 1947

Table 1. (Continued)

Species	Method[a]	Chemical	LC50 or EC50 (µg/L)[aa]	Species Mean Acute Value[aa] (µg/L)	Reference
Common carp, Cyprinus carpio	R, U	2-Methoxy ethyl mercuric chloride	139	139	Das & Misra, 1982
Fathead minnow, Pimephales promelas	S, M	Mercuric acetate	40	40	Curtis, et al. 1979; Curtis & Ward, 1981
Fathead minnow, Pimephales promelas	S, M	Mercuric thiocyanate	115	115	Curtis, et al. 1979; Curtis & Ward, 1981
Channel catfish (juvenile), Ictalurus punctatus	S, U	Ethylmercuric p-toluene sulfonanilide	51	51	Clemens & Sneed, 1959
Channel catfish (juvenile), Ictalurus punctatus	S, U	Ethylmercuric phosphate	49	49	Clemens & Sneed, 1959
Channel catfish (juvenile), Ictalurus punctatus	S, U	Phenylmercuric acetate	1,966	1,966	Clemens & Sneed, 1959
Channel catfish (juvenile), Ictalurus punctatus	S, U	Phenylmercuric acetate	28	28	Clemens & Sneed, 1958a, 1959
Channel catfish (juvenile), Ictalurus punctatus	S, U	Pyridylmercuric acetate	<176	-	Clemens & Sneed, 1958b
Channel catfish (juvenile), Ictalurus punctatus	S, U	Pyridylmercuric acetate	224	-	Clemens & Sneed, 1958b
Channel catfish (juvenile), Ictalurus punctatus	S, U	Pyridylmercuric acetate	<153	<182	Clemens & Sneed, 1958b
Mosquitofish (female), Gambusia affinis	S, U	Methoxy ethyl mercuric chloride	910	910	Joshi & Rege, 1980
Mosquitofish (female), Gambusia affinis	S, U	Phenylmercuric acetate	37	37	Joshi & Rege, 1980
Mosquitofish (female), Gambusia affinis	S, U	Phenylmercuric acetate (Ceresan)	44	44	Joshi & Rege, 1980

Table 1. (Continued)

Species	Method*	Chemical	LC50 or EC50 (µg/L)**	Species Mean Acute Value** (µg/L)	Reference
		SALTWATER SPECIES			
		Mercury(II)			
Polychaete worm (adult), *Neanthes arenaceodentata*	S, U	Mercuric chloride	96	-	Reish, et al. 1976
Polychaete worm (juvenile), *Neanthes arenaceodentata*	S, U	Mercuric chloride	100	97.98	Reish, et al. 1976
Sand worm (adult), *Nereis virens*	S, U	Mercuric chloride	70	70	Eisler & Hennekey, 1977
Polychaete worm (larva), *Capitella capitata*	S, U	Mercuric chloride	14	14	Reish, et al. 1976
Oligochaete worm, *Limnodriloides verrucosus*	R, U	Mercuric chloride	120	120	Chapman, et al. 1982a
Oligochaete worm, *Monopylephorus cuticulatus*	R, U	Mercuric chloride	230	230	Chapman, et al. 1982a
Oligochaete worm, *Tubificoides gabrielliae*	R, U	Mercuric chloride	98	98	Chapman, et al. 1982a
Northern horse mussel, *Modiolus modiolus*	S, M	Mercuric chloride	230	230	Hilmy, et al. 1981
Blue mussel, *Mytilus edulis*	S, U	Mercuric chloride	5.8	5.8	Martin, et al. 1981
Bay scallop (juvenile), *Argopecten irradians*	S, U	Mercuric chloride	89	89	Nelson, et al. 1976
Pacific oyster, *Crassostrea gigas*	S, U	Mercuric chloride	6.7	-	Martin, et al. 1981
Pacific oyster, *Crassostrea gigas*	S, M	Mercuric chloride	5.7	-	Glickstein, 1978
Pacific oyster, *Crassostrea gigas*	S, M	Mercuric nitrate	5.5	5.944	Glickstein, 1978

Table 1. (Continued)

Species	Method[a]	Chemical	LC50 or EC50 (µg/L)[**]	Species Mean Acute Value (µg/L)[**]	Reference
Eastern oyster, Crassostrea virginica	S, U	Mercuric chloride	5.6	-	Calabrese & Nelson, 1974; Calabrese, et al. 1977
Eastern oyster, Crassostrea virginica	S, U	Mercuric chloride	10.2	7.558	MacInnes & Calabrese, 1978
Common rangia (adult), Rangia cuneata	S, U	Mercuric chloride	10,000	-	Olson & Harrel, 1973
Common rangia (adult), Rangia cuneata	S, U	Mercuric chloride	8,700	-	Olson & Harrel, 1973
Common rangia (adult), Rangia cuneata	S, M	Mercuric chloride	58	-	Dillon, 1977
Common rangia (adult), Rangia cuneata	S, M	Mercuric chloride	122	*****	Dillon, 1977
Quahog clam, Mercenaria mercenaria	S, U	Mercuric chloride	4.8	4.8	Calabrese & Nelson, 1974; Calabrese, et al. 1977
Soft-shell clam (adult), Mya arenaria	S, U	Mercuric chloride	400	400	Eisler & Hennekey, 1977
Copepod, Pseudodiaptomus coronatus	S, U	Mercuric chloride	79	79	Gentile, 1982
Copepod, Eurytemora affinis	S, U	Mercuric chloride	158	158	Gentile, 1982
Copepod, Acartia clausi	S, U	Mercuric chloride	10	10	Gentile, 1982
Copepod (adult), Acartia tonsa	S, U	Mercuric chloride	10	-	Sosnowski & Gentile, 1978
Copepod (adult), Acartia tonsa	S, U	Mercuric chloride	14	-	Sosnowski & Gentile, 1978
Copepod (adult), Acartia tonsa	S, U	Mercuric chloride	15	-	Sosnowski & Gentile, 1978

Table 1. (Continued)

Species	Method[a]	Chemical	LC50 or EC50 (µg/L)[aa]	Species Mean Acute Value (µg/L)[aa]	Reference
Copepod (adult), *Acartia tonsa*	S, U	Mercuric chloride	20	14.32	Gentile, 1982
Copepod, *Nitocra spinipes*	S, U	Mercuric chloride	230	230	Bengtsson, 1978
Mysid, *Mysidopsis bahia*	FT, M	Mercuric chloride	3.5	3.5	Gentile, et al. 1982, 1983; Lussier, et al. Manuscript
White shrimp (adult), *Penaeus setiferus*	S, U	Mercuric chloride	17	17	Green, et al. 1976
American lobster (larva), *Homarus americanus*	S, U	Mercuric chloride	20	20	Johnson & Gentile, 1979
Hermit crab (adult), *Pagurus longicarpus*	S, U	Mercuric chloride	50	50	Eisler & Hennekey, 1977
Dungeness crab (larva), *Cancer magister*	S, U	Mercuric chloride	8.2	-	Martin, et al. 1981
Dungeness crab (larva), *Cancer magister*	S, M	Mercuric chloride	6.6	7.357	Glickstein, 1978
Green crab (larva), *Carcinus maenas*	S, U	Mercuric chloride	14	14	Connor, 1972
Starfish (adult), *Asterias forbesi*	S, U	Mercuric chloride	60	60	Eisler & Hennekey, 1977
Haddock (larva), *Melanogrammus aeglefinus*	S, U	Mercuric chloride	98	98	Cardin, 1982
Mummichog, *Fundulus heteroclitus*	S, U	Mercuric chloride	300	-	Dorfman, 1977
Mummichog, *Fundulus heteroclitus*	S, U	Mercuric chloride	200	-	Dorfman, 1977
Mummichog, *Fundulus heteroclitus*	S, U	Mercuric chloride	300	-	Dorfman, 1977

Table 1. (Continued)

Species	Method[a]	Chemical	LC50 or EC50 (µg/L)[**]	Species Mean Acute Value (µg/L)[**]	Reference
Mummichog, *Fundulus heteroclitus*	S, U	Mercuric chloride	300	-	Dorfman, 1977
Mummichog (adult), *Fundulus heteroclitus*	S, U	Mercuric chloride	800	-	Eisler & Hennekey, 1977
Mummichog (adult), *Fundulus heteroclitus*	S, U	Mercuric chloride	2,000	*****	Klaunig, et al. 1975
Mummichog (embryo), *Fundulus heteroclitus*	S, M	Mercuric chloride	67.4	67.4	Sharp & Neff, 1982
Atlantic silverside (larva), *Menidia menidia*	S, U	Mercuric chloride	144	-	Cardin, 1982
Atlantic silverside (larva), *Menidia menidia*	S, U	Mercuric chloride	125	-	Cardin, 1982
Atlantic silverside (juvenile), *Menidia menidia*	S, U	Mercuric chloride	86	115.7	Cardin, 1982
Tidewater silverside (juvenile), *Menidia peninsulae*	S, U	Mercuric chloride	71	71	Hansen, 1983
Fourspine stickleback (adult), *Apeltes quadracus*	S, U	Mercuric chloride	315	315	Cardin, 1982
Spot (juvenile), *Leiostomus xanthurus*	S, U	Mercuric chloride	36	36	Hansen, 1983
Winter flounder (larva), *Pseudopleuronectes americanus*	S, U	Mercuric chloride	1,820	-	Cardin, 1982
Winter flounder (larva), *Pseudopleuronectes americanus*	S, U	Mercuric chloride	1,560	-	Cardin, 1982

Table 1. (Continued)

Species	Method*	Chemical	LC50 or EC50 (µg/L)**	Species Mean Acute Value (µg/L)**	Reference
Winter flounder (larva), *Pseudopleuronectes americanus*	S, U	Mercuric chloride	1,810	-	Cardin, 1982
Winter flounder (larva), *Pseudopleuronectes americanus*	S, U	Mercuric chloride	1,320	-	Cardin, 1982
Winter flounder (larva), *Pseudopleuronectes americanus*	S, U	Mercuric chloride	1,960	1,678	Cardin, 1982
		Methylmercury			
Amphipod (adult), *Gammarus duebeni*	S, U	Methylmercuric chloride	150	150	Lockwood & Inman, 1975
Mummichog (embryo), *Fundulus heteroclitus*	S, M	Methylmercuric chloride	51.1	51.1	Sharp & Neff, 1982
		Other Mercury Compounds			
Grass shrimp (adult), *Palaemonetes pugio*	S, M	Mercuric acetate	47	47	Curtis, et al. 1979; Curtis & Ward, 1981
Grass shrimp (adult), *Palaemonetes pugio*	S, M	Mercuric thiocyanate	76	76	Curtis, et al. 1979; Curtis & Ward, 1981
Mummichog, *Fundulus heteroclitus*	S, U	Mercurous sulfate	6,800	-	Dorfman, 1977
Mummichog, *Fundulus heteroclitus*	S, U	Mercurous sulfate	300	*****	Dorfman, 1977

* S = static, R = renewal, FT = flow-through, U = unmeasured, M = measured.

** Results are expressed as mercury, not as the chemical.

*** Not used in calculation of Species Mean Acute Value because data are available for a more sensitive life stage.

**** "Less than" values were not used in calculations.

*****No Species Mean Acute Value calculated because acute values are too divergent for this species.

Table 2. Chronic Toxicity of Mercury to Aquatic Animals

Species	Test*	Chemical	Limits (µg/L)**	Chronic Value (µg/L)**	Reference
		FRESHWATER SPECIES			
		Mercury(II)			
Cladoceran, Daphnia magna	LC***	Mercuric chloride	0.72-1.28	0.96	Biesinger, et al. 1982
Cladoceran, Daphnia magna	LC****	Mercuric chloride	0.91-1.82	1.287	Biesinger, et al. 1982
Fathead minnow, Pimephales promelas	LC	Mercuric chloride	<0.26*****	<0.26	Snarski & Olson, 1982
Fathead minnow, Pimephales promelas	ELS	Mercuric chloride	<0.23*****	<0.23	Call, et al. 1983
		Methylmercury			
Cladoceran, Daphnia magna	LC***	Methylmercuric chloride	<0.04*****	<0.04	Biesinger, et al. 1982
Cladoceran, Daphnia magna	LC****	Methylmercuric chloride	0.52-0.87	0.6726	Biesinger, et al. 1982
Brook trout, Salvelinus fontinalis	LC	Methylmercuric chloride	0.29-0.93	0.5193	McKim, et al. 1976.
		Other Mercury Compounds			
Cladoceran, Daphnia magna	LC****	Phenylmercuric acetate	1.12-1.90	1.459	Biesinger, et al. 1982
		SALTWATER SPECIES			
		Mercury(II)			
Mysid, Mysidopsis bahia	LC	Mercuric chloride	0.8-1.6	1.131	Gentile, et al. 1982, 1983; Lussier, et al. Manuscript

Table 2. (Continued)

* LC = life cycle or partial life cycle, ELS = early life stage.

** Results are expressed as mercury, not as the chemical.

*** Flow-through

**** Renewal

*****Adverse effects occurred at this concentration, which was the lowest concentration tested.

Acute-Chronic Ratio

Species	Acute Value (µg/L)	Chronic Value (µg/L)	Ratio
	Mercury(II)		
Cladoceran, *Daphnia magna*	5	0.96	5.208
Cladoceran, *Daphnia magna*	5	1.287	3.885
Fathead minnow, *Pimephales promelas*	168	<0.26	>646.2
Fathead minnow, *Pimephales promelas*	150	<0.23	>652.2
Mysid, *Mysidopsis bahia*	3.5	1.131	3.095
	Methylmercury		
Brook trout *Salvelinus fontinalis*	73.89†	0.5193	142.3

† Geometric mean of 2 values from McKim, et al. (1976) in Table 1.

Table 3. Ranked Genus Mean Acute Values with Species Mean Acute-Chronic Ratios

Rank[a]	Genus Mean Acute Value (µg/L)	Species	Species Mean Acute Value (µg/L)	Species Mean Acute-Chronic Ratio
		FRESHWATER SPECIES		
		Mercury(II)		
28	2,000	Stonefly, *Acroneuria lycorias*	2,000	-
27	2,000	Mayfly, *Ephemerella subvaria*	2,000	-
26	2,000	Caddisfly, *Hydropsyche betteni*	2,000	-
25	1,200	Caddisfly, (Unidentified)	1,200	-
24	1,200	Damselfly, (Unidentified)	1,200	-
23	1,000	Worm, *Nais* sp.	1,000	-
22	1,000	Mozambique tilapia, *Tilapia mossambica*	1,000	-
21	406.2	Tubificid worm, *Spirosperma ferox*	330	-
		Tubificid worm, *Spirosperma nikolskyi*	500	-
20	370	Snail, *Aplexa hypnorum*	370	-
19	275	Rainbow trout, *Salmo gairdneri*	275	-
18	250	Tubificid worm, *Quistadrilus multisetosus*	250	-
17	240	Tubificid worm, *Rhyacodrilus montana*	240	-

Table 3. (Continued)

Rank*	Genus Mean Acute Value (µg/L)	Species	Species Mean Acute Value (µg/L)	Species Mean Acute-Chronic Ratio
16	240	Coho salmon, *Oncorhynchus kisutch*	240	-
15	180	Tubificid worm, *Limnodrilus hoffmeisteri*	180	-
14	180	Mosquitofish, *Gambusia affinis*	180	-
13	160	Bluegill, *Lepomis macrochirus*	160	-
12	158.7	Fathead minnow, *Pimephales promelas*	158.7	>649.2**
11	140	Tubificid worm, *Tubifex tubifex*	140	-
10	140	Tubificid worm, *Stylodrilus heringianus*	140	-
9	100	Tubificid worm, *Varichaeta pacifica*	100	-
8	80	Tubificid worm, *Branchiura sowerbyi*	80	-
7	80	Snail, *Amnicola* sp.	80	-
6	50	Crayfish, *Orconectes limosus*	50	-
5	30	Guppy, *Poecilia reticulata*	30	-
4	20	Crayfish, *Faxonella clypeatus*	20	-
3	20	Midge, *Chironomus* sp.	20	-

Table 3. (Continued)

Rank*	Genus Mean Acute Value (µg/L)	Species	Species Mean Acute Value (µg/L)	Species Mean Acute-Chronic Ratio
2	10	Amphipod, *Gammarus* sp.	10	-
1	2,646	Cladoceran, *Daphnia magna*	3,157	4.498**
		Cladoceran, *Daphnia pulex*	2,217	-
		SALTWATER SPECIES		
		Mercury(II)		
29	1,678	Winter flounder, *Pseudopleuronectes americanus*	1,678	-
28	400	Soft-shell clam, *Mya arenaria*	400	-
27	315	Fourspine stickleback, *Apeltes quadracus*	315	-
26	230	Northern horse mussel, *Modiolus modiolus*	230	-
25	230	Copepod, *Nitocra spinipes*	230	-
24	230	Oligochaete worm, *Monophylephorus cuticulatus*	230	-
23	158	Copepod, *Eurytemora affinis*	158	-
22	120	Oligochaete worm, *Limnodrilloides verrucosus*	120	-

Table 3. (Continued)

Rank[a]	Genus Mean Acute Value (µg/L)	Species	Species Mean Acute Value (µg/L)	Species Mean Acute-Chronic Ratio
21	98	Oligochaete worm, *Tubificoides gabrielae*	98	-
20	98	Haddock, *Melanogrammus aeglefinus*	98	-
19	97.98	Polychaete worm, *Neanthes arenaceodentata*	97.98	-
18	90.63	Atlantic silverside, *Menidia menidia*	115.7	-
		Tidewater silverside, *Menidia peninsulae*	71	-
17	89	Bay scallop, *Argopecten irradians*	89	-
16	79	Copepod, *Pseudodiaptomus coronatus*	79	-
15	70	Sand worm, *Nereis virens*	70	-
14	67.4	Mummichog, *Fundulus heteroclitus*	67.4	-
13	60	Starfish, *Arterias forbesi*	60	-
12	50	Hermit crab, *Pagurus longicarpus*	50	-
11	36	Spot, *Leiostomus xanthurus*	36	-
10	20	American lobster, *Homarus americanus*	20	-
9	17	White shrimp, *Penaeus setiferus*	17	-

Table 3. (Continued)

Rank*	Genus Mean Acute Value (µg/L)	Species	Species Mean Acute Value (µg/L)	Species Mean Acute-Chronic Ratio
8	14	Green crab, *Carcinus maenas*	14	-
7	14	Polychaete worm, *Capitella capitata*	14	-
6	11.97	Copepod, *Acartia clausi*	10	-
		Copepod, *Acartia tonsa*	14.32	
5	7.357	Dungeness crab, *Cancer magister*	7.357	-
4	6.703	Pacific oyster, *Crassostrea gigas*	5.944	-
		Eastern oyster, *Crassostrea virginica*	7.558	-
3	5.8	Blue mussel, *Mytilus edulis*	5.8	-
2	4.8	Quahog clam, *Mercenaria mercenaria*	4.8	-
1	3.5	Mysid, *Mysidopsis bahia*	3.5	3.095

* Ranked from most resistant to most sensitive based on Genus Mean Acute Value.

**Geometric mean of two values in Table 2.

Table 3. (Continued)

Mercury(II)

Final Acute-Chronic Ratio = 3.731 (see text)

Fresh water

Final Acute Value = 4.857 µg/L

Criterion maximum concentration = (4.857 µg/L) / 2 = 2.428 µg/L

Final Chronic Value = (4.857 µg/L) / 3.731 = 1.302 µg/L

Salt water

Final Acute Value = 4.125 µg/L

Criterion maximum concentration = (4.125 µg/L) / 2 = 2.062 µg/L

Final Chronic Value = (4.125 µg/L) / 3.731 = 1.106 µg/L

Table 4. Toxicity of Mercury to Aquatic Plants

Species	Chemical	Effect	Result (µg/L)[a]	Reference
		FRESHWATER SPECIES		
		Mercury(II)		
Alga, *Chlorella vulgaris*	Mercuric chloride	33-day EC50 (cell division inhibition)	1,030	Rosko & Rachlin, 1977
Alga, *Chlorella vulgaris*	Mercuric chloride	LC50	100-1,000	Gipps & Biro, 1978
Alga, *Chlorella vulgaris*	Mercuric chloride	LC50	148-296	Rai, 1979; Rai, et al. 1981
Alga, *Chlorella vulgaris*	Mercuric chloride	15-day EC50 (growth)	443-592	Rai, et al. 1981
Alga, *Anabaena flos-aquae*	Mercuric chloride	EC50 (growth)	53	Thomas & Montes, 1978
Blue alga, *Microcystis aeruginosa*	Mercuric chloride	8-day incipient inhibition	5	Bringmann, 1975; Bringmann & Kuhn, 1976, 1978a,b
Green alga, *Scenedesmus quadricauda*	Mercuric chloride	8-day incipient inhibition	70	Bringmann, 1975; Bringmann & Kuhn, 1976, 1978a,b, 1979, 1980b
Alga, *Selenastrum capricornutum*	Mercuric chloride	Inhibited growth	59	Slooff, et al. 1983
Eurasian watermilfoil, *Myriophyllum spicatum*	Mercuric chloride	32-day EC50 (root weight)	3,400	Stanley, 1974
		Methylmercury		
Alga, *Anabaena flos-aquae*	Methylmercuric chloride	EC50 (growth)	6.0	Thomas & Montes, 1978
Alga, *Chlorella vulgaris*	Methylmercuric chloride	15-day EC50 (growth)	0.8-4.0	Rai, et al. 1981

Table 4. (Continued)

Species	Chemical	Effect	Result (µg/L)*	Reference
		Other Mercury Compounds		
Alga, *Anabaena flos-aquae*	Phenylmercuric acetate	EC50 (growth)	2.8	Thomas & Montes, 1978
		SALTWATER SPECIES		
		Mercury(II)		
Alga, *Thalassiosira aestevalis*	Mercuric chloride	Reduced chlorophyll a	10	Hollibaugh, et al. 1980
Seaweed, *Ascophyllum nodosum*	Mercuric chloride	10-day EC50 (growth)	100	Stromgren, 1980
Diatom, *Ditylum brightwellii*	Mercuric chloride	5-day EC50 (growth)	10	Canterford & Canterford, 1980
Seaweed, *Fucus serratus*	Mercuric chloride	10-day EC50 (growth)	160	Stromgren, 1980
Seaweed, *Fucus spiralis*	Mercuric chloride	10-day EC50 (growth)	80	Stromgren, 1980
Seaweed, *Fucus vesiculosus*	Mercuric chloride	10-day EC50 (growth)	45	Stromgren, 1980
Giant kelp, *Macrocystis pyrifera*	Mercuric chloride	4-day EC50 (growth)	50	Clendenning & North, 1959
Seaweed, *Pelvetia canaliculata*	Mercuric chloride	10-day EC50 (growth)	130	Stromgren, 1980

* Results are expressed as mercury, not as the chemical.

Table 5. Bioaccumulation of Mercury by Aquatic Organisms

Species	Tissue	Chemical	Duration (days)	Bioconcentration Factor*	Reference
		FRESHWATER SPECIES			
		Mercury(II)			
Rainbow trout, Salmo gairdneri	Whole body	Mercuric chloride	60	1,800	Boudou & Ribeyre, 1984
Fathead minnow, Pimephales promelas	Whole body	Mercuric chloride	287	4,994**	Snarski & Olson, 1982
		Methylmercury			
Rainbow trout, Salmo gairdneri	Whole body	Methylmercuric chloride	60	11,000	Boudou & Ribeyre, 1984
Rainbow trout, Salmo gairdneri	Whole body	Methylmercuric chloride	75	85,700	Niimi & Lowe-Jinde, 1984
Brook trout, Salvelinus fontinalis	Muscle	Methylmercuric chloride	273	11,000-33,000	McKim, et al. 1976
Brook trout, Salvelinus fontinalis	Whole body	Methylmercuric chloride	273	10,000-23,000	McKim, et al. 1976
Brook trout, Salvelinus fontinalis	Muscle and whole body	Methylmercuric chloride	756	12,000	McKim, et al. 1976
Fathead minnow, Pimephales promelas	Whole body	Methylmercuric chloride	336	44,130-81,670	Olson, et al. 1975
		SALTWATER SPECIES			
		Mercury(II)			
Eastern oyster (adult), Crassostrea virginica	Soft parts	Mercuric chloride	74	10,000	Koptler, 1974
American lobster (adult), Homarus americanus	Tail muscle	Mercuric chloride	30	129	Thurberg, et al. 1977

Table 5. (Continued)

Species	Tissue	Chemical	Duration (days)	Bioconcentration Factor*	Reference
		Methylmercury			
Eastern oyster (adult), Crassostrea virginica	Soft parts	Methylmercuric chloride	74	40,000	Kopfler, 1974
		Other Mercury Compounds			
Eastern oyster (adult), Crassostrea virginica	Soft parts	Phenylmercuric chloride	74	40,000	Kopfler, 1974

* Results are based on mercury, not the chemical.

**From concentrations that caused adverse effects in a life-cycle test.

Maximum Permissible Tissue Concentration			
Consumer	Action Level or Effect	Concentration (mg/kg)	Reference
Man	Action level for edible fish or shellfish	1.0	U.S. FDA, 1984a,b
Mink, Mustela vison	Histological evidence of injury	≤1.1	Wobeser, 1976a,b
Brook trout, Salvelinus fontinalis	Death (700 days)	5-7	McKim, et al. 1976

Methylmercury

Freshwater Final Residue Value = (1.0 mg/kg) / 81,700 = 0.000012 mg/kg = 0.012 µg/L (see text)

Saltwater Final Residue Value = (1.0 mg/kg) / 40,000 = 0.000025 mg/kg = 0.025 µg/L

Mercury(II)

Freshwater Final Residue Value = (1.0 mg/kg) / 4,994 = 0.00020 mg/kg = 0.20 µg/L (see text)

Saltwater Final Residue Value = (1.0 mg/kg) / 10,000 = 0.00010 mg/kg = 0.10 µg/L (see text)

Table 6. Other Data on Effects of Mercury on Aquatic Organisms

Species	Chemical	Duration	Effect	Result (µg/L)*	Reference
		FRESHWATER SPECIES			
		Mercury(II)			
Alga, (Spring assemblages, predominantely diatoms)	Mercuric chloride	2 hrs	EC50 (reduced photosynthesis)	80	Blinn, et al. 1977
Alga, Ankistrodesmus braunii	Mercuric chloride	168-240 hrs	EC50 (inhibited lipid biosynthesis)	2,590	Matson, et al. 1972
Alga, Ankistrodesmus braunii	Mercuric chloride	24 days	Inhibited growth	74	Trevors, 1982
Alga, Ankistrodesmus sp.	Mercuric chloride	10 days	More toxic at pH = 5 than pH = 7	5	Baker, et al. 1983
Alga, Synedra ulna	Mercuric chloride	0.29 days	BCF=29,000	-	Fujita & Hashizume, 1972
Green alga, Scenedesmus quadricauda	Mercuric chloride	96 hrs	Incipient inhibition	30**	Bringmann & Kuhn, 1959a,b
Green alga, Scenedesmus quadricauda	Mercuric cyanide	96 hrs	Incipient inhibition	150**	Bringmann & Kuhn, 1959a,b
Bacteria, Escherichia coli	Mercuric chloride	-	Incipient inhibition	200	Bringmann & Kuhn, 1959a
Bacteria, Escherichia coli	Mercuric cyanide	-	Incipient inhibition	200	Bringmann & Kuhn, 1959a
Bacteria, Pseudomonas putida	Mercuric chloride	16 hrs	Incipient inhibition	10	Bringmann & Kuhn, 1976, 1977a, 1979, 1980b
Protozoan, Entosiphon sulcatum	Mercuric chloride	72 hrs	Incipient inhibition	18	Bringmann, 1978; Bringmann & Kuhn, 1979, 1980b, 1981
Protozoan, Chilomonas paramecium	Mercuric chloride	48 hrs	Incipient inhibition	15	Bringmann, et al. 1980; Bringmann & Kuhn, 1981
Protozoan, Uronema parduczi	Mercuric chloride	20 hrs	Incipient inhibition	67	Bringmann & Kuhn, 1980a, 1981

Table 6. (Continued)

Species	Chemical	Duration	Effect	Result (µg/L)*	Reference
Protozoan, *Microregma heterostoma*	Mercuric chloride	28 hrs	Incipient inhibition	150	Bringmann & Kuhn, 1959b
Protozoan, *Microregma heterostoma*	Mercuric cyanide	28 hrs	Incipient inhibition	160	Bringmann & Kuhn, 1959b
Hydra, *Hydra oligactis*	Mercuric chloride	48 hrs	LC50	56	Slooff, 1983; Slooff, et al. 1983
Planarian, *Dugesia lugubris*	Mercuric chloride	48 hrs	LC50	55	Slooff, 1983
Tubificid worm, *Tubifex tubifex*	Mercuric chloride	48 hrs	LC50	3,200	Qureshi, et al. 1980
Snail, *Lymnaea stagnalis*	Mercuric chloride	48 hrs	LC50	443	Slooff, 1983; Slooff, et al. 1983
Mussel, *Margaritifera margaritifera*	Mercuric nitrate	39 days	BCF=302	-	Mellinger, 1973
Cladoceran, *Diaphanosoma sp.*	Mercuric chloride	3 wks	Reduced population density	2.8	Marshall, et al. 1981
Cladoceran, *Daphnia galeata mendotae*	Mercuric chloride	3 wks	Reduced population density	2.2	Marshall, et al. 1981
Cladoceran, *Daphnia magna*	Mercuric chloride	3 wks	Reproductive impairment	3.4	Biesinger & Christensen, 1972
Cladoceran, *Daphnia magna*	Mercuric chloride	48 hrs	EC50	30**	Bringmann & Kuhn, 1959a,b
Cladoceran, *Daphnia magna*	Mercuric cyanide	48 hrs	EC50	20**	Bringmann & Kuhn, 1959a,b
Cladoceran, *Daphnia magna*	Mercuric chloride	24 hrs	LC50	13	Bringmann & Kuhn, 1977b
Cladoceran, *Bosmina longirostris*	Mercuric chloride	3 wks	Reduced population density	2.8	Marshall, et al. 1981

Table 6. (Continued)

Species	Chemical	Duration	Effect	Result (µg/L)[a]	Reference
Natural copepod assemblages	Mercuric chloride	7 days	Reduced growth rate	28.3	Borgmann, 1980
Amphipod, *Gammarus* sp.	Mercuric chloride	7 days	BCF=2,500	-	Zubarik & O'Connor, 1978
Amphipod, *Gammarus* sp.	Mercuric nitrate	7 days	BCF=2,500	-	Zubarik & O'Connor, 1978
Crayfish (male, mixed ages), *Faxonella clypeatus*	Mercuric chloride	72 hrs	LC50	200	Holt & Fingerman, 1977
Crayfish (0.2 g), *Faxonella clypeatus*	Mercuric chloride	24 hrs	LC50	1,000	Holt & Fingerman, 1977
Crayfish (1.2 g), *Faxonella clypeatus*	Mercuric chloride	672 hrs	LC50	1,000	Holt & Fingerman, 1977
Crayfish (adult), *Orconectes limosus*	Mercuric chloride	96 hrs	LC60	740	Doyle, et al. 1976
Crayfish (juvenile), *Orconectes limosus*	Mercuric chloride	30 days	LC50 (unfed)	2	Boutet & Chalsemartin, 1973
Crayfish (juvenile), *Orconectes limosus*	Mercuric chloride	30 days	LC50 (fed)	<2	Boutet & Chalsemartin, 1973
Crayfish (male, mixed ages), *Procambarus clarki*	Mercuric chloride	72 hrs	LC50	200	Holt & Fingerman, 1977
Freshwater community (primary producers, herbivores and carnivorous midges)	Mercuric chloride	1 yr	Reduced algal standing stock and diversity; no evidence of effects on midges	0.1	Sigmon, et al. 1977
Mosquito, *Aedes aegypti*	Mercuric chloride	48 hrs	LC50	4,100	Slooff, et al. 1983
Mosquito, *Aedes aegypti*	Mercuric chloride	48 hrs	LC50	776	Slooff, et al. 1983

Table 6. (Continued)

Species	Chemical	Duration	Effect	Result (µg/L)*	Reference
Pink salmon (embryo), Oncorhynchus gorbuscha	Mercuric sulfate	2 days <time from fertilization to hatch	EC32 to EC81 (deformity)	5.2	Servizi & Martens, 1978
Pink salmon (pre-eyed embryo), Oncorhynchus gorbuscha	Mercuric sulfate	2 days <time from fertilization to stage	LC100	8.5	Servizi & Martens, 1978
Pink salmon (larva), Oncorhynchus gorbuscha	Mercuric sulfate	168 hrs	LC50	140	Servizi & Martens, 1978
Sockeye salmon (embryo), Oncorhynchus nerka	Mercuric sulfate	2 days <time from fertilization to hatch	EC45.6 (deformity)	4.3	Servizi & Martens, 1978
Sockeye salmon (pre-eyed embryo), Oncorhynchus nerka	Mercuric sulfate	2 days <time from fertilization to stage	LC100	9.3	Servizi & Martens, 1978
Sockeye salmon (larva), Oncorhynchus nerka	Mercuric sulfate	168 hrs	LC50	290	Servizi & Martens, 1978
Sockeye salmon (juvenile), Oncorhynchus nerka	Mercuric sulfate	168 hrs	LC50	190	Servizi & Martens, 1978
Rainbow trout (juvenile), Salmo gairdneri	Mercuric chloride	24 hrs	LC50	903	Wobeser, 1973
Rainbow trout (juvenile), Salmo gairdneri	Mercuric chloride	2 hrs	Depressed olfactory bulber response	74	Hara, et al. 1976
Rainbow trout (juvenile), Salmo gairdneri	Mercuric chloride	-	Growth inhibition	2.1-21	Matida, et al. 1971
Rainbow trout, Salmo gairdneri	Mercuric chloride	1 wk	Effected osmo-regulation	100	Lock, et al. 1981
Rainbow trout, Salmo gairdneri	Mercuric chloride	80 min	Avoidance threshold	0.2	Black & Birge, 1980
Rainbow trout (embryo, larva), Salmo gairdneri	Mercuric chloride	28 days	EC50 (death and deformity)	4.7 5.0	Birge, et al. 1979, 1980

Table 6. (Continued)

Species	Chemical	Duration	Effect	Result (µg/L)[a]	Reference
Rainbow trout (embryo, larva), *Salmo gairdneri*	Mercuric chloride	28 days	EC50 (death and deformity)	<0.1	Birge, et al. 1979, 1980
Rainbow trout (embryo, larva), *Salmo gairdneri*	Mercuric chloride	28 days	EC10 (death and deformity)	0.9	Birge, et al. 1981
Rainbow trout, *Salmo gairdneri*	Mercuric chloride	5, 18 mo	Substantial mortality	0.12-0.24	Birge, et al. 1979
Rainbow trout, *Salmo gairdneri*	Mercuric chloride	48 hrs	LC50	480	Slooff, et al. 1983
Goldfish (embryo, larva), *Carassius auratus*	Mercuric chloride	7 days	EC50 (death and deformity)	121.9	Birge, 1978; Birge, et al. 1979
Goldfish (embryo, larva), *Carassius auratus*	Mercuric chloride	7 days	EC50 (death and deformity)	0.7	Birge, 1978; Birge, et al. 1979
Common carp, *Cyprinus carpio*	Mercuric chloride	60-72 hrs	Reduced hatching success	≥3,000	Huckabee & Griffith, 1974
Fathead minnow, *Pimephales promelas*	Mercuric chloride	48 hrs	LC50	37	Slooff, et al. 1983
White sucker (adult), *Catostomus commersoni*	Mercuric chloride	36 min	Blood enzyme (LDH) inhibition 20%	8,000	Christensen, 1972
White sucker (adult), *Catostomus commersoni*	Mercuric chloride	46 min	Blood enzyme (GOT) inhibition 20%	10,000	Christensen, 1972
Channel catfish (embryo, larva), *Ictalurus punctatus*	Mercuric chloride	10 days	EC50 (death and deformity)	30	Birge, et al. 1979
Channel catfish (embryo, larva), *Ictalurus punctatus*	Mercuric chloride	10 days	EC50 (death and deformity)	0.3	Birge, et al. 1979
Channel catfish (embryo, larva), *Ictalurus punctatus*	Mercuric chloride	-	Increased albinism	0.5	Westerman & Birge, 1978
Channel catfish (embryo, larva), *Ictalurus punctatus*	Mercuric chloride	10 days	BCF=441-2071	-	Birge, et al. 1979

Table 6. (Continued)

Species	Chemical	Duration	Effect	Result (µg/L)[a]	Reference
Channel catfish (embryo, larva), Ictalurus punctatus	Mercuric chloride	10 days	BCF-4.4-353	-	Birge, et al. 1979
Mosquitofish, Gambusia affinis	Mercuric chloride	>10 days	LC50	500	Boudou, et al. 1979
Guppy, Poecilia reticulata	Mercuric chloride	24 hrs	LC50	13	Hamdy, 1977
Guppy, Poecilia reticulata	Mercuric chloride	48 hrs	LC50	303	Slooff, et al. 1983
Bluegill (embryo, larva), Lepomis macrochirus	Mercuric chloride	7-8 days	EC50 (death and deformity)	88.7	Birge, et al. 1979
Redear sunfish (embryo, larva), Lepomis microlophus	Mercuric chloride	7-8 days	EC50 (death and deformity)	137.2	Birge, et al. 1979
Largemouth bass (embryo, larva), Micropterus salmoides	Mercuric chloride	8 days	EC50 (death and deformity)	130 140	Birge, et al. 1978, 1979
Largemouth bass (embryo, larva), Micropterus salmoides	Mercuric chloride	8 days	EC50 (death and deformity)	5.3	Birge, et al. 1979
Largemouth bass, Micropterus salmoides	-	24 hrs	Affected opercular rhythm	10	Morgan, 1979
Mozambique tilapia, Tilapia mossambica	Mercuric nitrate	35 days	Clinical symptoms	310	Panigrahi & Misra, 1980
Mozambique tilapia, Tilapia mossambica	Mercuric chloride	48 hrs	LC50	1,000	Menezes & Qasim, 1983
Pig frog (embryo, larva), Rana grylio	Mercuric chloride	7 days	EC50 (death and deformity)	67.2	Birge, et al. 1979
River frog (embryo, larva), Rana heckscheri	Mercuric chloride	7 days	EC50 (death and deformity)	59.9	Birge, et al. 1979
Leopard frog (embryo, larva), Rana pipiens	Mercuric chloride	7 days	EC50 (death and deformity)	7.3	Birge, et al. 1979

Table 6. (Continued)

Species	Chemical	Duration	Effect	Result (µg/L)[a]	Reference
Narrow-mouthed toad (embryo, larva), Gastrophryne carolinensis	Mercuric chloride	7 days	EC50 (death and deformity)	1 1.3	Birge, et al. 1978, 1979
Green toad (embryo, larva), Bufo debilis	Mercuric chloride	7 days	EC50 (death and deformity)	40.0	Birge, et al. 1979
Fowler's toad (embryo, larva), Bufo fowleri	Mercuric chloride	7 days	EC50 (death and deformity)	65.9	Birge, et al. 1979
Red-spotted toad (embryo, larva), Bufo punctatus	Mercuric chloride	7 days	EC50 (death and deformity)	36.8	Birge, et al. 1979
Northern cricket frog (embryo, larva), Acris crepitans	Mercuric chloride	7 days	EC50 (death and deformity)	10.4	Birge, et al. 1979
Southern gray treefrog (embryo, larva), Hyla chrysoscelis	Mercuric chloride	7 days	EC50 (death and deformity)	2.4	Birge, et al. 1979
Spring peeper (embryo, larva), Hyla crucifer	Mercuric chloride	7 days	EC50 (death and deformity)	2.8	Birge, et al. 1979
Barking treefrog (embryo, larva), Hyla gratiosa	Mercuric chloride	7 days	EC50 (death and deformity)	2.5	Birge, et al. 1979
Squirrel treefrog (embryo, larva), Hyla squirella	Mercuric chloride	7 days	EC50 (death and deformity)	2.4	Birge, et al. 1979
Gray treefrog (embryo, larva), Hyla versicolor	Mercuric chloride	7 days	EC50 (death and deformity)	2.6	Birge, et al. 1979
African clawed frog, Xenopus laevis	Mercuric chloride	11 mos	Substantial mortality	0.16-0.2	Birge, et al. 1978
African clawed frog, Xenopus laevis	Mercuric chloride	48 hrs	LC50	74	Slooff & Baerselman, 1980; Slooff, et al. 1983

Table 6. (Continued)

Species	Chemical	Duration	Effect	Result (µg/L)[a]	Reference
Marbled salamander (embryo, larva), Ambystoma opacum	Mercuric chloride	7-8 days	EC50 (death and deformity)	108 107.5	Birge, et al. 1978, 1979
		Methylmercury			
Alga, Ankistrodesmus braunii	Methylmercuric chloride	168-240 hrs	Lipid biosynthesis, >EC50	1,598	Matson, et al. 1972
Alga, Coelastrum microporum	Methylmercuric chloride	-	EC50 (growth inhibition)	>2.4-<4.8	Holderness, et al. 1975
Alga, Scenedesmus obliquus	Methylmercuric chloride	14 days	BCF=2,100 (Maximum by third day)***	-	Havlik, et al. 1979
Alga, Microcystis incerta	Methylmercuric chloride	14 days	BCF=990 (Maximum by third day)***	-	Havlik, et al. 1979
Planarian, Dugesia dorotocephala	Methylmercuric chloride	4 days	LC50	200-500	Best, et al. 1981
Mussel, Margaritifera margaritifera	Methylmercuric chloride	57 days	BCF=2,463	-	Mellinger, 1973
Amphipod, Gammarus sp.	Methylmercuric chloride	7 days	BCF=8,000 (approx.)	-	Zubarik & O'Connor, 1978
Rainbow trout (juvenile), Salmo gairdneri	Methylmercuric chloride	84 days****	BCF=4,530 (whole fish, 5 C)	-	Reinert, et al. 1974
Rainbow trout (juvenile), Salmo gairdneri	Methylmercuric chloride	84 days****	BCF=6,620 (whole fish, 10 C)	-	Reinert, et al. 1974
Rainbow trout (juvenile), Salmo gairdneri	Methylmercuric chloride	84 days****	BCF=8,049 (whole fish, 15 C)	-	Reinert, et al. 1974
Rainbow trout, Salmo gairdneri	Methylmercuric chloride	-	inhibited growth	0.0037-0.037	Matida, et al. 1971
Rainbow trout, Salmo gairdneri	Methylmercuric chloride	14 days	approximate LC80	8	Blanc, 1973

Table 6. (Continued)

Species	Chemical	Duration	Effect	Result (µg/L)[a]	Reference
Rainbow trout, *Salmo gairdneri*	Methylmercuric chloride	1 wk	Effected osmo-regulation	5	Lock, et al. 1981
Rainbow trout, *Salmo gairdneri*	Methylmercuric chloride plus inorganic mercury	120 days	Loss of appetite	48 mg/kg in food	Matida, et al. 1971
Rainbow trout (embryo, larva), *Salmo gairdneri*	-	24 days	EC50 (death and deformity)	5	Birge & Black, 1977
Rainbow trout, *Salmo gairdneri*	Methylmercuric chloride plus inorganic mercury	269 days	Loss of nervous control	48 mg/kg in food	Matida, et al. 1971
Rainbow trout, *Salmo gairdneri*	Methylmercuric chloride	30 min	EC50 (Reduced viability of sperm)	1,000	McIntyre, 1973
Brook trout (embryo), *Salvelinus fontinalis*	Methylmercuric chloride	16-17 days	Decreased enzyme (GOT) activity	0.88	Christensen, 1975
Brook trout (alevin), *Salvelinus fontinalis*	Methylmercuric chloride	Incubation period + 21 days	Reduced growth	0.7	Christensen, 1975
Brook trout (alevins), *Salvelinus fontinalis*	Methylmercuric chloride	30 days	Increased enzyme (GOT) activity	0.79	Christensen, 1975
Brook trout (juvenile), *Salvelinus fontinalis*	Methylmercuric chloride	14 days	Increased blood plasma chloride	2.93	Christensen, et al. 1977
Brook trout, *Salvelinus fontinalis*	Methylmercuric chloride	8 days	Increased cough frequency	≥3	Drummond, et al. 1974
Common carp, *Cyprinus carpio*	Methylmercuric chloride	16 days	Reduced protein synthesis	0.05	Sharma & Davis, 1980b
Mosquitofish, *Gambusia affinis*	Methylmercuric chloride	<24 hrs	LC50	500	Boudou, et al. 1979

Table 6. (Continued)

Species	Chemical	Duration	Effect	Result (µg/L)[a]	Reference
Mosquitofish, *Gambusia affinis*	Methylmercuric chloride	30 days	BCF=2,500 (whole fish, 10 C)	-	Boudou, et al. 1979
Mosquitofish, *Gambusia affinis*	Methylmercuric chloride	30 days	BCF=4,300 (whole fish, 18 C)	-	Boudou, et al. 1979
Mosquitofish, *Gambusia affinis*	Methylmercuric chloride	30 days	BCF=3,000 (whole fish, 164 mg/kg in food, 10 C)	-	Boudou, et al. 1979
Mosquitofish, *Gambusia affinis*	Methylmercuric chloride	30 days	BCF=27,000 (whole fish, 238 ng/kg in food, 26 C)	-	Boudou, et al. 1979
Bluegill (juvenile), *Lepomis macrochirus*	Methylmercuric chloride	28.5 days	BCF=373***** (whole fish, 9 C)	-	Camber, et al. 1978
Bluegill (juvenile), *Lepomis macrochirus*	Methylmercuric chloride	28.5 days	BCF=921***** (whole fish, 21 C)	-	Camber, et al. 1978
Bluegill (juvenile), *Lepomis macrochirus*	Methylmercuric chloride	28.5 days	BCF=2,400***** (whole fish, 33 C)	-	Camber, et al. 1978
Leopard frog (tadpole), *Rana pipiens*	Methylmercuric chloride	48 hrs	LC100	50-100	Chang, et al. 1974
Leopard frog, *Rana pipiens*	Methylmercuric chloride	4 mos	Failure to metamorphose	1-10	Chang, et al. 1974
Leopard frog (blastula embryo), *Rana pipiens*	Methylmercuric chloride	5 days	LC50	12-16	Dial, 1976

Table 6. (Continued)

Species	Chemical	Duration	Effect	Result (µg/L)[a]	Reference
Leopard frog (gastrula embryo), Rana pipiens	Methylmercuric chloride	5 days	LC50	8-12	Dial, 1976
Leopard frog (neural plate embryo), Rana pipiens	Methylmercuric chloride	5 days	LC50	12-16	Dial, 1976
Leopard frog (blastula embryo), Rana pipiens	Methylmercuric chloride	96 hrs	EC50 (teratogenesis)	0-4	Dial, 1976
Leopard frog (gastrula embryo), Rana pipiens	Methylmercuric chloride	96 hrs	EC50 (teratogenesis)	8-12	Dial, 1976
Leopard frog (neural plate embryo), Rana pipiens	Methylmercuric chloride	96 hrs	EC50 (teratogenesis)	12	Dial, 1976
Newt, Triturus viridescens	Methylmercuric chloride	≥2 days	Delayed limb regeneration	8	Chang, et al. 1976
Newt, Triturus viridescens	Methylmercuric chloride	17 days	Death	300	Chang, et al. 1976
Newt, Triturus viridescens	Methylmercuric chloride	8 days	Death	1,000	Chang, et al. 1976
Mink (adult), Mustela vison	Methylmercuric chloride	93 days	Histologic evidence of injury	1,100	Wobeser, 1973
Mink (adult), Mustela vison	Methylmercuric chloride	93 days	LC50 in brain tissue	11,000	Wobeser, 1973
		Other Mercury Compounds			
Alga, (Florida Lake assemblage)	Methylmercuric dicyandiamide	125 hrs	Reduced biomass	0.8 (approx.)	Harriss, et al. 1970

Table 6. (Continued)

Species	Chemical	Duration	Effect	Result (µg/L)[a]	Reference
Alga, (Florida Lake assemblage)	N-Methylmercuric-1,2,3,6-tetrahydro-3,6-methano-3,4,5,6,7,7,-hexachloro-phthalimide	125 hrs	Reduced biomass	0.3 (approx.)	Harriss, et al. 1970
Alga, Cladophoraceae	Ethylmercuric phosphate	1 hr	Nuisance control	38.6	Burrows & Combs, 1958
Alga, Ulothrichaceae	Ethylmercuric phosphate	1 hr	Nuisance control	38.6	Burrows & Combs, 1958
Alga, (Florida Lake assemblage)	Phenylmercuric acetate	125 hrs	Reduced biomass	0.5 (approx.)	Harriss, et al. 1970
Alga, (Florida Lake assemblage)	Diphenyl mercury	125 hrs	Reduced biomass	2.8 (approx.)	Harriss, et al. 1970
Alga, Scenedesmus obliquus	Phenylmercuric chloride	14 days	BCF=13,000 (Maximum by third day)	-	Havlik, et al. 1979
Alga, Microcystis incerta	Phenylmercuric chloride	14 days	BCF=4,000 (Maximum by third day)	-	Havlik, et al. 1979
Sponge, Ephydatia fluviatilis	Mercury	30 days	Malformed gemmoscleres	1	Mysing-Gubala & Poirrier, 1981
Sponge, Ephydatia fluviatilis	Mercury	30 days	LC50	100-500	Mysing-Gubala & Poirrier, 1981
Amphipod, Gammarus sp.	Phenylmercuric acetate	7 days	BCF=8,000 (approx.)	-	Zubarik & O'Connor, 1978
Crayfish (juvenile), Procambarus clarki	Methylmercuric dicyandimide	120 hrs	LC50	56	Hendrick & Everett, 1965
Sockeye salmon (juvenile), Oncorhynchus nerka	Pyridylmercuric acetate	1.5 hrs	LC50	10,500-15,700	Burrows & Palmer, 1949
Sockeye salmon (juvenile), Oncorhynchus nerka	Pyridylmercuric acetate	1 hr	Safe for disease control	<954	Rucker, 1948

Table 6. (Continued)

Species	Chemical	Duration	Effect	Result (µg/L)[a]	Reference
Sockeye salmon (juvenile), Oncorhynchus nerka	Pyridylmercuric acetate	1 hr	Safe for disease control	<4,752	Rucker & Whipple, 1951
Chinook salmon (fingerling), Oncorhynchus tshawytscha	Ethylmercuric phosphate	1 hr	Distress	77	Burrows & Combs, 1958
Chinook salmon, Oncorhynchus tshawytscha	Ethylmercuric phopshate	20 hrs	Safe for disease control	39	Burrows & Combs, 1958
Rainbow trout (juvenile), Salmo gairdneri	Pyridylmercuric acetate	1 hr	LC100	1,030	Allison, 1957
Rainbow trout (juvenile), Salmo gairdneri	Pyridylmercuric acetate	1 hr	LC0	967	Allison, 1957
Rainbow trout (juvenile), Salmo gairdneri	Pyridylmercuric acetate	1 hr	LC33 (8.3 C) (13.3 C)	4,750 4,750	Rodgers, et al. 1951
Rainbow trout (alevin), Salmo gairdneri	Pyridylmercuric acetate	1 hr	Safe for disease control	<4,750	Rucker & Whipple, 1951
Rainbow trout (juvenile), Salmo gairdneri	Pyridylmercuric acetate	1 hr	LC60	517	Allison, 1957
Rainbow trout, Salmo gairdneri	Phenylmercuric acetate	12 wks	Growth inhibition	0.11-1.1	Matida, et al. 1971
Rainbow trout (juvenile), Salmo gairdneri	Ethylmercuric phosphate	48 hrs	LC50	43	Matida, et al. 1971
Rainbow trout (juvenile), Salmo gairdneri	Ethylmercuric p-toluene sulfonanilide	-	Retarded learning	5 µg/g in feed daily or 10 µg/g feed every fifth day	Hartman, 1978
Rainbow trout (juvenile), Salmo gairdneri	Pyridylmercuric acetate	24 hrs	LC50	25	MacLeod & Pessah, 1973
Rainbow trout (juvenile), Salmo gairdneri	Phenylmercuric acetate	48 hrs	LC50	1,780	Williford, 1966

Table 6. (Continued)

Species	Chemical	Duration	Effect	Result (µg/L)*	Reference
Rainbow trout (juvenile), Salmo gairdneri	Merthiolate	48 hrs	LC50	10,500	Wilford, 1966
Brown trout (juvenile), Salmo trutta	Pyridylmercuric acetate	1 hr	Safe for disease control	4,750	Rodgers, et al. 1951
Brown trout (juvenile), Salmo trutta	Pyridylmercuric acetate	48 hrs	LC50	2,950	Wilford, 1966
Brown trout (juvenile), Salmo trutta	Merthiolate	48 hrs	LC50	26,800	Wilford, 1966
Brook trout (juvenile), Salvelinus fontinalis	Pyridylmercuric acetate	1 hr	Safe for disease control	2,070	Allison, 1957
Brook trout (juvenile), Salvelinus fontinalis	Pyridylmercuric acetate	1 hr	Safe for disease control	4,750	Rodgers, et al. 1951
Brook trout (juvenile), Salvelinus fontinalis	Pyridylmercuric acetate	48 hrs	LC50	5,080	Wilford, 1966
Brook trout (juvenile), Salvelinus fontinalis	Merthiolate	48 hrs	LC50	36,900	Wilford, 1966
Lake trout (juvenile), Salvelinus namaycush	Pyridylmercuric acetate	48 hrs	LC50	3,610	Wilford, 1966
Lake trout (juvenile), Salvelinus namaycush	Merthiolate	48 hrs	LC50	1,060	Wilford, 1966
Channel catfish (juvenile), Ictalurus punctatus	Pyridylmercuric acetate	72 hrs	LC50	232	Clemens & Sneed, 1958a, 1959
Channel catfish (juvenile), Ictalurus punctatus	Pyridylmercuric acetate	48 hrs	LC50 (10 C) (16.5 C) (24 C)	1,960 1,340 234	Clemens & Sneed, 1958b
Channel catfish (yolk sac fry), Ictalurus punctatus	Pyridylmercuric acetate	48 hrs	LC50 (23 C)	178	Clemens & Sneed, 1958b
Channel catfish (1 wk-old), Ictalurus punctatus	Pyridylmercuric acetate	48 hrs	LC50 (23 C)	<148	Clemens & Sneed, 1958b

Table 6. (Continued)

Species	Chemical	Duration	Effect	Result (µg/L)[a]	Reference
Channel catfish, Ictalurus punctatus	Pyridylmercuric acetate	48 hrs	LC50	1,370	Williford, 1966
Channel catfish, Ictalurus punctatus	Merthiolate	48 hrs	LC50	2,800	Williford, 1966
Bluegill (juvenile), Lepomis macrochirus	Pyridylmercuric acetate	48 hrs	LC50	7,600	Williford, 1966
Bluegill (juvenile), Lepomis macrochirus	Merthiolate	48 hrs	LC50	32,000	Williford, 1966
Largemouth bass, Micropterus salmoides	Mercury	21 days	Threshold of effect opercular rhythm	10	Morgan, 1979
SALTWATER SPECIES					
	Mercury(II)				
Red alga, Antithamnion plumula	Mercuric chloride	30 min	LC50 after 7 days	5,000	Boney & Corner, 1959
Alga, Chaetoceros galvestonensis	Mercuric chloride	4 days	About 30% reduction in growth	10	Hannan, et al. 1973b
Alga, Chaetoceros galvestonensis	Mercuric chloride	4 days	No growth of culture	100	Hannan, et al. 1973b
Alga, Chaetoceros galvestonensis	Mercuric chloride	4 days	BCF=10,920	-	Hannan, et al. 1973b
Alga, Chroomonas salina	Mercuric chloride	2 days	BCF=853	-	Parrish & Carr, 1976
Alga, Cyclotella sp.	Mercuric chloride	3 days	No growth of culture	100	Hannan & Patouillet, 1972
Alga, Dunaliella sp.	Mercuric chloride	-	75% reduction in CO_2	2,500	Mills & Colwell, 1977

Table 6. (Continued)

Species	Chemical	Duration	Effect	Result (µg/L)[a]	Reference
Alga, *Dunaliella tertiolecta*	Mercuric chloride	8 days	About 10% increase in maximum chlorophyll *a* concentration	100	Betz, 1977
Alga, *Dunaliella tertiolecta*	Mercuric chloride	8 days	About 45% increase in maximum chlorophyll *a* concentration	220	Betz, 1977
Diatom, *Nitzchia acicularis*	Mercuric chloride	7 days	Prevented growth	150-200	Mora & Fabregas, 1980
Diatom, *Skeletonema costatum*	Mercuric chloride	15 days	Reduced cell density	0.08	Cloutier-Mantha & Harrison, 1980
Alga, *Dunaliella tertiolecta*	Mercuric chloride	3 days	About 15% reduction in growth	10	Davies, 1976
Alga, *Dunaliella tertiolecta*	Mercuric chloride	8 days	No effect on growth	2	Davies, 1976
Alga, *Isochrysis galbana*	Mercuric chloride	15 days	About 10% reduction in growth	5.1	Davies, 1974
Alga, *Isochrysis galbana*	Mercuric chloride	15 days	About 60% reduction in growth	10.5	Davies, 1974
Alga, *Isochrysis galbana*	Mercuric chloride	28 days	Growth rate recovery to near normal after day 5	10.5	Davies, 1974
Kelp (zoospores, gametophytes, sporophytes), *Laminaria hyperborea*	Mercuric chloride	28 days	Lowest concentration causing growth inhibition	10	Hopkins & Kain, 1971
Kelp (zoospores, gametophytes, sporophytes), *Laminaria hyperborea*	Mercuric chloride	22 hrs	EC50 respiration	about 450	Hopkins & Kain, 1971
Kelp (zoospores, gametophytes, sporophytes), *Laminaria hyperborea*	Mercuric chloride	28 hrs	About 80% reduction in respiration	10,000	Hopkins & Kain, 1971

Table 6. (Continued)

Species	Chemical	Duration	Effect	Result (µg/L)*	Reference
Alga, *Phaeodactylum tricornutum*	Mercuric chloride	4 days	About 50% reduction in growth	50	Hannan, et al. 1973b
Alga, *Phaeodactylum tricornutum*	Mercuric chloride	4 days	No growth of culture	120	Hannan, et al. 1973a
Alga, *Phaeodactylum tricornutum*	Mercuric chloride	4 days	BCF=7,120	-	Hannan, et al. 1973b
Red alga (sporling), *Plumaria elegans*	Mercuric chloride	24 hrs	40% reduction in growth over 21 days	120	Boney, 1971
Red alga (sporling), *Plumaria elegans*	Mercuric chloride	1 hr	40% reduction in growth over 21 days	1,000	Boney, 1971
Red alga (sporling), *Plumaria elegans*	Mercuric chloride	18 hrs	LC50 after 7 days	3,170	Boney, et al. 1959
Red alga, *Plumaria elegans*	Mercuric chloride	30 min	LC50 after 7 days	6,700	Boney & Corner, 1959
Red alga, *Polysiphonia lanosa*	Mercuric chloride	30 min	LC50 after 7 days	8,000	Boney & Corner, 1959
Alga (mixed), *Asterionella japonica* plus *Diogenes* sp.	Mercuric chloride	8 days	BCF=3,467	-	Laumond, et al. 1973
5 seaweed species, *Ascophyllum nodosum*, *Fucus spiralis*, *F. versiculosus*, *F. serratus*, *Pelvetia canaliculata*	Mercuric chloride	10 days	10-30% reduction in growth	10	Stromgren, 1980
Algae, (eighteen species)	Mercuric chloride	17 days	Growth inhibition	<5-15	Berland, et al. 1976

Table 6. (Continued)

Species	Chemical	Duration	Effect	Result (µg/l)*	Reference
Algae, (eighteen species)	Mercuric chloride	17 days	Death	10-50	Berland, et al. 1976
Algae, (three species)	Mercuric chloride	168 hrs	Depressed growth	30-350	Sick & Windom, 1975
Algae, (three species)	Mercuric chloride	168 hrs	No further bioaccumulation	40	Sick & Windom, 1975
Algae, (three species)	Mercuric chloride	168 hrs	Changes in cell chemistry	30-350	Sick & Windom, 1975
Natural phytoplankton populations	Mercuric chloride	120 hrs	Reduced chlorophyll a	6	Hollibaugh, et al. 1980
Natural phytoplankton populations	Mercuric chloride	96 hrs	Reduced biomass	2	Hollibaugh, et al. 1980
Phytoplankton, (Natural assemblages)	Mercuric chloride	21 days	Inhibited growth	1	Thomas, et al. 1977
Protozoan, *Cristigera* sp.	Mercuric chloride	12 hrs	Reduced growth	2.5-5	Gray & Ventilla, 1973
Protozoan, *Euplotes vannus*	Mercuric chloride	48 hrs	Inhibition of reproduction	1,000	Persoone & Uyttersprot, 1975
Sand worm (adult), *Nereis virens*	Mercuric chloride	168 hrs	LC50	60	Eisler & Hennekey, 1977
Sand worm (adult), *Nereis virens*	Mercuric chloride	168 hrs	LC100	125	Eisler & Hennekey, 1977
Polychaete worm (adult), *Ophryotrocha diadema*	Mercuric chloride	96 hrs	LC13	50	Reish & Carr, 1978
Polychaete worm (adult), *Ophryotrocha diadema*	Mercuric chloride	96 hrs	LC60	100	Reish & Carr, 1978
Polychaete worm (adult), *Ophryotrocha diadema*	Mercuric chloride	96 hrs	LC100	500	Reish & Carr, 1978
Polychaete worm, *Ophryotrocha diadema*	Mercuric chloride	48 hrs	LC50	30-100	Parker, 1984

Table 6. (Continued)

Species	Chemical	Duration	Effect	Result (µg/L)*	Reference
Blue mussel (larva), *Mytilus edulis*	Mercuric chloride	24 hrs	Abnormal development	32	Okubo & Okubo, 1962
Pacific oyster (larva), *Crassostrea gigas*	Mercuric chloride	24 hrs	Abnormal development	32	Okubo & Okubo, 1962
Eastern oyster (embryo), *Crassostrea virginica*	Mercuric chloride	12 days	LC50	12	Calabrese & Nelson, 1974; Calabrese, et al. 1977
Eastern oyster (embryo), *Crassostrea virginica*	Mercuric chloride	48 hrs	LC0	1	Calabrese, et al. 1973
Eastern oyster (embryo), *Crassostrea virginica*	Mercuric chloride	19 days	Trace metal upset	50	Kopfler, 1974
Clam, *Mulina lateralis*	Mercuric chloride	72 hrs	Reduced calcium uptake	26.5	Ming-Shan & Zubkoff, 1982
Common rangia, *Rangia cuneata*	Mercuric chloride	96 hrs	LC50 (<1 g/kg salinity)	5,100	Olson & Harrel, 1973
Common rangia, *Rangia cuneata*	Mercuric chloride	14 days	BCF=1,130 (whole animal)	-	Dillon & Neff, 1978
Quahog clam (larva), *Mercenaria mercenaria*	Mercuric chloride	8-10 days	LC50	14	Calabrese & Nelson, 1974; Calabrese, et al. 1977
Quahog clam (larva), *Mercenaria mercenaria*	Mercuric chloride	42-48 hrs	LC0	2.5	Calabrese, et al. 1973
Soft-shell clam (adult), *Mya arenaria*	Mercuric chloride	168 hrs	LC0	1	Elsler & Hennekey, 1977
Soft-shell clam (adult), *Mya arenaria*	Mercuric chloride	168 hrs	LC50	4	Elsler & Hennekey, 1977
Soft-shell clam (adult), *Mya arenaria*	Mercuric chloride	168 hrs	LC100	30	Elsler & Hennekey, 1977
Copepods (adult), (5 genera)	Mercuric chloride	10 days	90% decrease in egg production	10	Reeve, et al. 1977
Copepods (adult), (5 genera)	Mercuric chloride	10 days	70% decrease in fecal pellet	10	Reeve, et al. 1977

Table 6. (Continued)

Species	Chemical	Duration	Effect	Result (µg/L)*	Reference
Copepods (adult), (5 genera)	Mercuric chloride	48 hrs	Hg-Cu interactions on LC50	17 (Hg in mixture)	Reeve, et al. 1977
Copepod (adult), Pseudocalanus minutus	Mercuric chloride	70 days	No growth of culture	5	Sonntag & Greve, 1977
Copepod (adult), Pseudocalanus minutus	Mercuric chloride	70 days	No growth inhibition	1	Sonntag & Greve, 1977
Copepod (adult), Acartia clausi	Mercuric chloride	1.9 hrs	LC50	50	Corner & Sparrow, 1956
Copepod (adult), Acartia clausi	Mercuric chloride	24 hrs	BCF=7,500	-	Reiichiro, et al. 1983
Barnacle (adult), Balanus balanoides	Mercuric chloride	48 hrs	LC90	1,000	Clarke, 1947
Barnacle (cyprid), Balanus balanoides	Mercuric chloride	6 hrs	About 10% reduciton in substrate attachment over 19 days	10	Pyefinch & Mott, 1948
Barnacle (cyprid), Balanus balanoides	Mercuric chloride	6 hrs	LC50	90	Pyefinch & Mott, 1948
Barnacles (naupilius), Balanus crenatus	Mercuric chloride	6 hrs	LC50	60	Pyefinch & Mott, 1948
Barnacle (cyprid), Balanus improvisus	Mercuric chloride	48 hrs	About 50% abnormal development	16,600	Clarke, 1947
White shrimp (adult), Penaeus setiferus	Mercuric chloride	60 days	No effect on respiration, growth, or molting	1	Green, et al. 1976
Grass shrimp (larva), Palaemonetes vulgaris	Mercuric chloride	<24 hrs	LC100	56	Shealy & Sandifer, 1975
Grass shrimp (larva), Palaemonetes vulgaris	Mercuric chloride	48 hrs	LC0	<5.6	Shealy & Sandifer, 1975
Grass shrimp, Palaemonetes pugio	Mercuric chloride	120 hrs	LC50	148	Barthalmus, 1977

Table 6. (Continued)

Species	Chemical	Duration	Effect	Result (µg/L)[a]	Reference
Grass shrimp, *Palaemonetes pugio*	Mercuric chloride	24 hrs	Impaired conditioned avoidance response	37	Barthalmus, 1977
Grass shrimp (larva), *Palaemonetes vulgaris*	Mercuric chloride	48 hrs	LC50	10	Shealy & Sandifer, 1975
Grass shrimp (larva), *Palaemonetes vulgaris*	Mercuric chloride	48 hrs	Abnormal development	10-18	Shealy & Sandifer, 1975
Hermit crab (adult), *Pagurus longicarpus*	Mercuric chloride	168 hrs	LC0	10	Elsier & Hennekey, 1977
Hermit crab (adult), *Pagurus longicarpus*	Mercuric chloride	168 hrs	LC50	50	Elsier & Hennekey, 1977
Hermit crab (adult), *Pagurus longicarpus*	Mercuric chloride	168 hrs	LC100	125	Elsier & Hennekey, 1977
Green crab (adult), *Carcinus maenas*	Mercuric chloride	48 hrs	LC50	1,000	Portmann, 1968
Green crab (adult), *Carcinus maenas*	Mercuric chloride	48 hrs	LC50	1,200	Connor, 1972
Green crab (larva), *Carcinus maenas*	Mercuric chloride	47 hrs	LC50	10	Connor, 1972
Green crab (larva), *Carcinus maenas*	Mercuric chloride	20-30 hrs	LC50	33	Connor, 1972
Green crab (larva), *Carcinus maenas*	Mercuric chloride	4.3-13.5 hrs	LC50	100	Connor, 1972
Green crab (larva), *Carcinus maenas*	Mercuric chloride	2.7 hrs	LC50	1,000	Connor, 1972
Green crab (larva), *Carcinus maenas*	Mercuric chloride	0.5 hrs	LC50	3,300	Connor, 1972
Green crab (larva), *Carcinus maenas*	Mercuric chloride	0.22 hrs	LC50	10,000	Connor, 1972

Table 6. (Continued)

Species	Chemical	Duration	Effect	Result (µg/L)*	Reference
Fiddler crab (adult), Uca pugilator	Mercuric chloride	28 days	Low survival, inhibited limb regeneration	1,000	Weis, 1976
Fiddler crab (adult), Uca pugilator	Mercuric chloride	6 days	20-25% reduction in percent survival	180	Vernberg & Vernberg, 1972
Fiddler crab (adult), Uca pugilator	Mercuric chloride	6 days	20-25% reduction in percent survival	180	Vernberg & Vernberg, 1972
Fiddler crab (adult), Uca pugilator	Mercuric chloride	24 hrs	Increased oxygen consumption	180	Vernberg & Vernberg, 1972
Fiddler crab (zoea), Uca pugilator	Mercuric chloride	8 days	LC50	1.8	DeCoursey & Vernberg, 1972
Fiddler crab (zoea), Uca pugilator	Mercuric chloride	24 hrs	20-100% increase in metabolic rate after stage I zoea	1.8	DeCoursey & Vernberg, 1972
Fiddler crab (zoea), Uca pugilator	Mercuric chloride	5 days	About 40% increase in swimming activity of stage V zoea	1.8	DeCoursey & Vernberg, 1972
Starfish (adult), Asterias forbesi	Mercuric chloride	168 hrs	LC0	10	Eisler & Hennekey, 1977
Starfish (adult), Asterias forbesi	Mercuric chloride	168 hrs	LC50	20	Eisler & Hennekey, 1977
Starfish (adult), Asterias forbesi	Mercuric chloride	168 hrs	LC100	125	Eisler & Hennekey, 1977
Sea urchin (spermatazoa), Arbacia punctulata	Mercuric chloride	8 min	About 150% increase in swimming speed	20	Young & Nelson, 1974
Sea urchin (spermatazoa), Arbacia punctulata	Mercuric chloride	24 min	About 80% decrease in swimming speed	2,000	Young & Nelson, 1974
Sea urchin (embryo), Arbacia punctulata	Mercuric chloride	13 hrs	Abnormal development	92	Waterman, 1937
Haddock (embryo), Melanogrammus aeglefinus	Mercuric chloride	96 hrs	LC50	918	Cardin, 1982

Table 6. (Continued)

Species	Chemical	Duration	Effect	Result (µg/L)[a]	Reference
Mummichog (adult), Fundulus heteroclitus	Mercuric chloride	168 hrs	LC0	100	Eisler & Hennekey, 1977
Mummichog (adult), Fundulus heteroclitus	Mercuric chloride	168 hrs	LC50	800	Eisler & Hennekey, 1977
Mummichog (adult), Fundulus heteroclitus	Mercuric chloride	168 hrs	LC100	1,000	Eisler & Hennekey, 1977
Mummichog (adult), Fundulus heteroclitus	Mercuric chloride	24 hrs	Disrupted osmoregulation	125	Renfro, et al. 1974
Mummichog (adult), Fundulus heteroclitus	-	96 hrs	Affected liver enzymes	200	Jacklm, et al. 1970
Mummichog (adult), Fundulus heteroclitus	Mercuric chloride	28 days	Up to 40% reduction in enzyme activity before recovery	12	Jacklm, 1973
Mummichog (embryo), Fundulus heteroclitus	Mercuric chloride	3 days	Many developmental abnormalities	30-40	Wels & Wels, 1977
Mummichog (embryo), Fundulus heteroclitus	Mercuric chloride	3 days	Some developmental abnormalities	10-20	Wels & Wels, 1977
Mummichog (embryo), Fundulus heteroclitus	Mercuric chloride	12 hrs	Some developmental abnormalities	30-40	Wels & Wels, 1977
Mummichog (embryo), Fundulus heteroclitus	Mercuric chloride	32 days	EC50	67.4	Sharp & Neff, 1980
Mummichog (larva), Fundulus heteroclitus	Mercuric chloride	-	No effect	50	Wels & Wels, 1983
Mummichog (adult), Fundulus heteroclitus	Mercuric chloride	-	Mercury redistribution organs following Se pretreatment	1,000 ug Hg/kg body wt plus 400 ug Se/kd body wt	Sheline & Schmidt-Nielson, 1977
Mummichog (adult), Fundulus heteroclitus	Mercuric chloride	96 hrs	Cellular degeneration	250-5,000	Gardner, 1975
Mummichog (adult), Fundulus heteroclitus	Mercuric chloride	48 hrs	LC100	2,000	Eisler, et al. 1972

Table 6. (Continued)

Species	Chemical	Duration	Effect	Result (µg/L)*	Reference
Mummichog (adult), *Fundulus heteroclitus*	Mercuric chloride	96 hrs	Sluggish, uncoordinated swimming	1,150	Klaunig, et al. 1975
Shiner perch, *Cymatogaster aggregata*	Mercuric chloride	-	45% reduction of brain cholinesterase activity	33,900	Abou-Donia & Menzel, 1967
Striped bass (adult), *Morone saxatilis*	Mercuric chloride	30 days	Decreased respiration 30 days post exposure	5	Dawson, et al. 1977
Winter flounder (adult), *Pseudopleuronectes americanus*	Mercuric chloride	60 days	Decreased respiration	10	Calabrese, et al. 1975
		Methylmercury			
Alga, *Dunaliella tertiolecta*	Methylmercuric chloride	10 min	EC50 (photosynthesis)	about 170	Overnell, 1975
Alga, *Phaeodactylum tricornutum*	Methylmercuric chloride	25 days	EC50 (photosynthesis)	about 190	Overnell, 1975
Red alga (sporling), *Plumaria elegans*	Methylmercuric chloride	18 hrs	LC50 after 7 days	44	Boney, et al. 1959
Alga, *Tetraselmis succica*	Methylmercuric chloride	3 days	Inhibited growth	25	Mora & Fabregas, 1980
Alga, *Chaetoceros* sp.	Dimethylmercury	3 days	About 75% reduction in growth	100	Hannan & Patoulliet, 1972
Alga, *Cyclotella* sp.	Dimethylmercury	3 days	About 15% reduction in growth	500	Hannan & Patoulliet, 1972
Alga, *Phaeodactylum* sp.	Dimethylmercury	3 days	About 45% reduction in growth	500	Hannan & Patoulliet, 1972
Red alga (sporling), *Plumaria elegans*	Methylmercuric chloride	25 min	EC50 (growth over 21 days	40	Boney, 1971
Diatom, *Nitzchia acicularis*	Methylmercuric chloride	3 days	Inhibited growth	25	Mora & Fabregas, 1980

Table 6. (Continued)

Species	Chemical	Duration	Effect	Result (µg/L)[a]	Reference
Dinoflagellate, *Scrippsiella faeroense*	Mercuric acetate	14 days	No growth of culture	1,000	Kayser, 1976
Alga, *Chlorella* sp.	Ethylmercuric phosphate	10 days	22% reduction in growth	0.6	Ukeles, 1962
Alga, *Chlorella* sp.	Ethylmercuric phosphate	10 days	100% lethal to culture	6	Ukeles, 1962
Alga, *Dunaliella euchlora*	Ethylmercuric phosphate	10 days	36% reduction in growth	0.6	Ukeles, 1962
Alga, *Dunaliella euchlora*	Ethylmercuric phosphate	10 days	100% lethal to culture	60	Ukeles, 1962
Alga, *Monochrysis lutheri*	Ethylmercuric phosphate	10 days	No reduction in growth	0.6	Ukeles, 1962
Alga, *Monochrysis lutheri*	Ethylmercuric phosphate	10 days	100% lethal to culture	6	Ukeles, 1962
Alga, *Phaeodactylum tricornutum*	Ethylmercuric phosphate	10 days	45% reduction in growth	0.6	Ukeles, 1962
Alga, *Phaeodactylum tricornutum*	Ethylmercuric phosphate	10 days	100% lethal to culture	6	Ukeles, 1962
Alga, *Protococcus* sp.	Ethylmercuric phosphate	10 days	14% reduction in growth	0.6	Ukeles, 1962
Alga, *Protococcus* sp.	Ethylmercuric phosphate	10 days	100% lethal to culture	6	Ukeles, 1962
Red alga (sporling), *Plumaria elegans*	Mercuric iodide	18 hrs	LC50 after 7 days	156	Boney, et al. 1959
Red alga (sporling), *Plumaria elegans*	Ethylmercuric chloride	18 hrs	LC50 after 7 days	26	Boney, et al. 1959
Red alga (sporling), *Plumaria elegans*	Phenylmercuric chloride	18 hrs	LC50 after 7 days	54	Boney, et al. 1959

Table 6. (Continued)

Species	Chemical	Duration	Effect	Result (µg/L)*	Reference
Diatom, *Nitzchia delicatissima*	Methylmercuric dicyandiamide	24 hrs	EC50 (photosynthesis)	0.4	Harriss, et al. 1970
Blue mussel (adult), *Mytilus edulis*	Methylmercuric chloride	24 hrs	About 90% reduced feeding rate	400	Dorn, 1976
Eastern oyster (adult), *Crassostrea virginica*	Methylmercuric chloride	19 days	Trace metal upset	50	Koptler, 1974
Copepod (adult), *Acartia clausi*	Methylmercuric chloride	24 hrs	BCF=56,000-350,000	-	Reichiro, et al. 1983
Amphipod (adult), *Gammarus duebeni*	Methylmercuric chloride	3 days	Induced diuresis	56	Lockwood & Inman, 1975
Fiddler crab (adult), *Uca* sp.	Methylmercuric chloride	32 days	No limb regeneration	300-500	Weis, 1977
Fiddler crab (adult), *Uca* sp.	Methylmercuric chloride	32 days	Melanin absent in regerated limbs	100	Weis, 1977
Mummichog (adult), *Fundulus heteroclitus*	Methylmercuric chloride	24 hrs	Disrupted osmoregulation	125	Renfro, et al. 1974
Mummichog (embryo), *Fundulus heteroclitus*	Methylmercuric chloride	7 days	Teratological effects	50	Weis, et al. 1981
Mummichog (larva), *Fundulus heteroclitus*	Methylmercuric chloride	-	Reduced LT50s	50	Weis & Weis, 1983
Mummichog (embryo), *Fundulus heteroclitus*	Methylmercuric chloride	-	Developed resistance in a pond	50	Weis & Weis, 1984
Striped mullet, *Mugil cephalus*	Methylmercuric chloride	13 days	Inhibited fin regeneration	1	Weis & Weis, 1978
			Other Mercury Compounds		
Dinoflagellate, *Gymnodinium spendens*	Mercuric acetate	11 days	55% reduction in growth	10	Kayser, 1976
Dinoflagellate, *Scrippsiella faeroense*	Mercuric acetate	25 days	45% reduction in growth, morphological variation	10	Kayser, 1976

Table 6. (Continued)

Species	Chemical	Duration	Effect	Result (µg/L)*	Reference
Red alga (sporling), Plumaria elegans	Phenylmercuric iodide	18 hrs	LC50 after 7 days	104	Boney, et al. 1959
Red alga (sporling), Plumaria elegans	Isoamylmercuric chloride	18 hrs	LC50 after 7 days	19	Boney, et al. 1959
Red alga (sporling), Plumaria elegans	n-amylmercuric chloride	18 hrs	LC50 after 7 days	13	Boney, et al. 1959
Red alga (sporling), Plumaria elegans	Isopropylmercuric chloride	18 hrs	LC50 after 7 days	28	Boney, et al. 1959
Red alga (sporling), Plumaria elegans	n-propylmercuric chloride	18 hrs	LC50 after 7 days	13	Boney, et al. 1959
Red alga (sporling), Plumaria elegans	n-butylmercuric chloride	18 hrs	LC50 after 7 days	13	Boney, et al. 1959
Diatom, Nitzchia delicatissima	N-methylmercuric-1,2,3,6-tetrahydro-3,6-methano-3,4,5,6,7,7-hexachloro-phthalimine	24 hrs	EC50 (photosynthesis)	0.3	Harriss, et al. 1970
Diatom, Nitzchia delicatissima	Phenylmercuric acetate	24 hrs	EC50 (photosynthesis)	1.5	Harriss, et al. 1970
Diatom, Nitzchia delicatissima	Diphenylmercury	24 hrs	EC50 (photosynthesis)	18	Harriss, et al. 1970
Diatom, Nitzchia acicularis	Phenylmercuric acetate	7 days	Inhibited growth	25	Mora & Fabregas, 1980
Eastern oyster (adult), Crassostrea virginica	Mercuric acetate	12 hrs daily for 15 days	33% reduction in shell growth	10	Cunningham, 1976
Eastern oyster (adult), Crassostrea virginica	Mercuric acetate	60 days	LC55	100	Cunningham, 1976
Eastern oyster (adult), Crassostrea virginica	Phenylmercuric chloride	19 days	Trace metal upset	50	Kopfler, 1974

Table 6. (Continued)

Species	Chemical	Duration	Effect	Result (µg/L)*	Reference
Copepod (adult), Acartia clausi	Mercuric acetate	1.9 hrs	LC50	50	Corner & Sparrow, 1956
Copepod (adult), Acartia clausi	Ethylmercuric chloride	1.9 hrs	LC50	50	Corner & Sparrow, 1956
Coho salmon (adult), Oncorhynchus kisutch	Pyridylmercuric acetate	12-15 wks, 1 hr wkly as juveniles	0.03 mg Hg/kg wet wt muscle 2 yrs post-exposure	1,000	Amend, 1970
Sockeye salmon (juvenile), Oncorhynchus nerka	Pyridylmercuric acetate	12-15 wks, 1 hr wkly	1.2 mg Hg/kg wet wt muscle 12 weeks post-exposure	1,000	Amend, 1970
Sockeye salmon (adult), Oncorhynchus nerka	Pyridylmercuric acetate	12-15 wks, 1 hr wkly as juveniles	0.24 mg Hg/kg wet wt muscle 3 yrs post-exposure	1,000	Amend, 1970
Sockeye salmon (adult), Oncorhynchus nerka	Pyridylmercuric acetate	12 1-hr exposures as juveniles	0.04 mg Hg/kg wet wt muscle 4 yrs post-exposure	1,000	Amend, 1970
Chinook salmon (adult), Oncorhynchus tshawytscha	Pyridylmercuric acetate	35 wks, 1 hr wkly as juveniles	up to 0.12 mg Hg/kg muscle 4 yrs later	1,000	Amend, 1970
Threespine stickleback, Gasterosteus aculeatus	Phenylmercuric acetate	370 min	LC100	100	Boetius, 1960

* Results are expressed as mercury, not as the chemical.

** In river water.

*** Static, continual loss over time.

**** Not at steady-state.

*****BCF independent of concentration in water over range tested.

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